# CYCLOPROPYLINDOLE DERIVATIVES

Technical Field

5   This invention relates to cyclopropylindoles (CPI) (which term is
used to encompass cyclopropylbenzindoles (CBI)) compounds and
their precursors, to methods of synthesizing these compounds on
solid supports, and to compounds of utility therein. This
invention further relates to collections of these compounds, and
10   methods for identifying and isolating. CPI and precursor compounds
with useful and diverse activities from such collections.

Background to the invention

15   A large number of both synthetic and naturally occurring low
molecular weight ligands are known that interact with DNA via a
number of different mechanisms, including covalent or non-covalent
interaction in the minor or major grooves, intercalation between
base pairs or other types of non-specific interactions.

20   Of the class of ligands which interact with the minor groove, GC
specific ligands include Chromomycin, pyrrolo[2,1-
c][1,4]benzodiazepines (PBDs), Mitomycins and Ecteinascidins. Of
these, all but Chromomycin form a covalent bond with the DNA. Of
25   AT specific ligands, cyclopropylindoles form covalent bonds,
whilst compounds such as distamycin and netropsin do not.

Cyclopropylindole (CPI) compounds are a class of highly potent
antitumour antibiotics which includes the natural products CC-1065
30   (V.L. Reynolds et al, J. Antibiot., 39, 1986, 319-314) and the
duocarmycins (D.L. Boger, Pure & Appl. Chem., 66, 1994, 837-844),
having IC_{50}s in the low pM range in tumour cells growing in $vitro$.
They are of the general structures A and B:

A   B
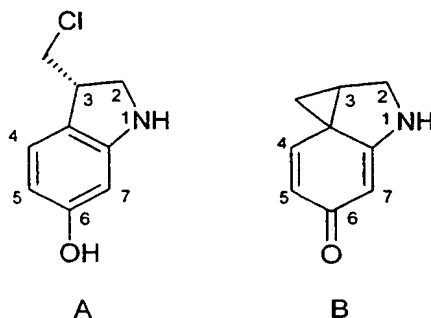
Studies with compounds that model the binding subunit have shown
5   that the more stable open chain seco-precursors (e.g. A) are as
potent as the cyclopropylindole compounds (e.g. B). Further, ring
closure is not essential for DNA binding.

A number of synthetic analogues of the natural products have been
10 prepared in which the oxygen of A is protected as a carbamate that
must be cleaved (by non-specific enzymatic hydrolysis) for
activity. Further analogues of a similar type are disclosed in
WO88/04659 and WO91/16324. Analogues where the 6 substituent is N
or S are disclosed in WO 97/07097 and WO 98/11101.

15 Compounds having biological activity can be identified by
screening collections of compounds (i.e. libraries of compounds)
produced through synthetic chemical techniques. Such screening
methods include methods wherein the library comprises a plurality
20 of compounds synthesized at specific locations on the surface of a
solid support where a receptor is appropriately labelled to
identify binding to the compound, e.g., fluorescent or radioactive
labels. Correlation of the labelled receptor bound to the support
with its location on the support identifies the binding compound
25 (US 5,143,854).

Central to these methods is the screening of a multiplicity of
compounds in the library and the ability to identify the
structures of the compounds which have a requisite biological
30 activity. In order to facilitate synthesis and identification,
the compounds in the library are typically formed on solid
supports. Usually each such compound is covalently attached to

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the support via a cleavable or non-cleavable linking arm. The
libraries of compounds can be screened either on the solid support
or as cleaved products to identify compounds having good
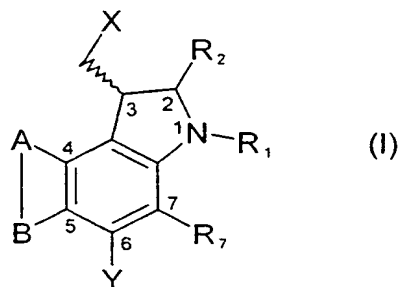biological activity.

Disclosure of the Invention

The present invention provides CPI compounds with structures that
allow them to be joined to combinatorial chains, as well as
combinatorial libraries containing CPIs themselves.

A first aspect of the present invention relates to compounds of
formula **I**:

(I)

wherein X is an electrophilic leaving group;

Y is selected from NH-Prot, O-Prot, S-Prot, NO_2, NHOH, N_3, NHR,
NRR, N=NR, N(O)RR, $NHSO_2R$, N=NPhR, SR or SSR, where Prot
represents a protecting group;

A and B collectively represent a fused benzene or pyrrole ring (in
either orientation), which is optionally substituted by up to
respectively 4 or 2 groups independently selected from R, OH, OR,
halo, nitro, amino, Me_3Sn, CO_2H, CO_2R;

R_1 is a nitrogen protecting group, where if Y includes a
protecting group, these protecting groups are orthogonal;

R_2 and R_7 are independently selected from H, R, OH, OR, halo,
nitro, amino, Me_3Sn;

wherein R is selected from

(a) a lower alkyl group having 1 to 10 carbon atoms,

(b) an aralkyl group (i.e. an alkyl group with one or more aryl
substituents), preferably of up to 12 carbon atoms;

the alkyl group of (a) or (b) optionally containing one or more carbon-carbon double or triple bonds, which may form part of a conjugated system; and

(c) an aryl group, preferably of up to 12 carbon atoms; and

5   wherein R is optionally substituted by one or more halo, hydroxy, amino, or nitro groups, and optionally contains one or more hetero atoms, which may form part of, or be, a functional group;

except that when R_1 is Boc, Y is NO_2, X is Cl, and R_2 and R_7 are H, then A and B do not collectively represent either an unsubstituted

10   benzene ring or:

$$MeO_2C \diagup \diagdown \underset{\underset{H}{N}}{}$$
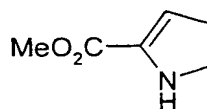
If R is an aryl group, and contains a hetero atom, then R is a

15   heterocyclic group. If R is an alkyl chain, and contains a hetero atom, the hetero atom may be located anywhere in the alkyl chain, e.g. $-O-C_2H_5$, $-CH_2-S-CH_3$, or may form part of, or be, a functional group, e.g. carbonyl, hydroxy.

20   R is preferably independently selected from a lower alkyl group having 1 to 10 carbon atoms, or an aralkyl group, preferably of up to 12 carbon atoms, or an aryl group, preferably of up to 12 carbon atoms, optionally substituted by one or more halo, hydroxy, amino, or nitro groups. It is more preferred that R is

25   independently selected from lower alkyl groups having 1 to 10 carbon atoms optionally substituted by one or more halo, hydroxy, amino, or nitro groups. It is particularly preferred that R is an unsubstituted straight or branched chain alkyl group, having 1 to 10, preferably 1 to 6, and more preferably 1 to 4, carbon atoms,

30   e.g. methyl, ethyl, propyl, butyl.

Y is preferably NH-Prot, O-Prot, S-Prot, and more preferably NH-Prot.

5

These compounds are useful in the synthesis of collections of CBI and CPI precursors. Compounds of formula **I**:

(I)
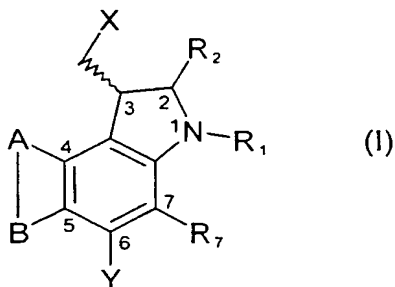
wherein X, A, B, R_1, R_2, R_7 are as defined above, and Y is selected from NH_2, NH-Prot, OH, O-Prot, SH, S-Prot, NO_2, NHOH, N_3, NHR, NRR,

5 N=NR, N(O)RR, $NHSO_2R$, N=NPhR, SR or SSR, where Prot represents a protecting group can be attached to a solid support, e.g. via a connecting link which may comprise a chain of combinatorial units. This is a second aspect of the invention, i.e. the use of these compounds in methods of combinatorial chemistry synthesis, wherein

10 the compound is joined to a solid support by a chain comprising at least one combinatorial unit. The preferences for R expressed in the first aspect apply to this aspect of the invention.

Y in this second aspect is preferably NH_2, NH-Prot, OH, O-Prot, SH,

15 S-Prot, more preferably NH_2, NH-Prot, SH, S-Prot, and most preferably NH-Prot. As an alternative, OH and O-Prot are preferred.

Furthermore, compounds of formula (I) with substituents as defined in this aspect, where the fused ring, represented by -A-B-, bears a

20 substituent $-CO_2H$ or $-CO_2R$, may be used as combinatorial units (see below).

The term 'protecting group' (and more specifically 'nitrogen protecting group') has the meaning usual in synthetic chemistry,

25 particularly for 'nitrogen protecting group', in synthetic peptide chemistry. It means any group which may be covalently bound to the protected atom of the CBI or CPI grouping, and permits reactions to be carried out upon the molecule containing this grouping without its removal. Nevertheless, it is able to be

30 removed from the protected atom without affecting the remainder of

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the molecule. Suitable nitrogen protecting groups for the present invention include Fmoc (9-fluorenylmethoxycarbonyl), Nvoc (6-nitroveratryloxycarbonyl), Teoc (2-trimethylsilylethyloxycarbonyl), Troc

5   (2,2,2-trichloroethyloxycarbonyl), Boc (t-butyloxycarbonyl), CBZ (benzyloxycarbonyl), Alloc (allyloxycarbonyl) or Psec (2(-phenylsulphonyl)ethyloxycarbonyl). Suitable oxygen (hydroxyl) protecting groups include t-butyl ethers, Benzyl ethers, Silyl ethers, MOM (methoxy methyl ethers), MEM (2-methoxy ethoxy methyl

10   ethers) or acetates. Suitable sulphur (thiol) protecting groups include benzyl, nitrogenzyl thioether or fluorenylmethyl thioethers. Other suitable groups are described in Protective Groups in Organic Synthesis, T Green and P Wuts, published by Wiley, 1991, which is incorporated herein by reference.

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In the first and second aspects of the invention it is preferred that the nitrogen protecting group has a carbamate functionality where it binds to the nitrogen atom to be protected.

20   The term 'orthogonal' in relation to 'protecting groups' has the meaning usual in synthetic chemistry. It means that one protecting group may be selectively removed without affecting the other protecting group. This is achieved by using protecting groups which are sensitive to different removal conditions.

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Examples of orthogonal protecting group pairs are:
CBZ carbamate and Boc Carbamate - the CBZ group can be removed by hydrogenation whilst the BOC carbamate remains intact. Alternatively, the BOC group can be removed with TFA whilst the

30   CBZ group remains intact. The same applies to a CBZ carbamate and a tert-butyl ether.

Fmoc carbamate and t-butyl ether - The Fmoc group can be removed with base (50% piperidine in DMF) without affecting the t-butyl

35   protection. The t-butyl side chain protecting group is removed by acid (TFA, trifluoroacetic acid). If necessary t-butyl ethers can be cleaved in the presence of Fmoc as Fmoc is unaffected by acid.

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Alloc carbamate and Silyl ethers - an allyl carbamate is cleaved
specifically by zero valent palladium catalysts, Silyl ethers are
cleaved with fluoride ions or dilute acid (neither reagent affects
Alloc and palladium does not affect Silyl ethers).

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An example of non-orthogonal pair includes CBZ and Fmoc, as both
are cleaved by hydrogenation.

An electrophilic leaving group is a group that is readily
10   eliminated from the molecule and carries with it an electron-pair.
These may be termed "nucleogufugal" leaning groups.

In the present invention, it is preferred that X is either halogen
or OSO_2R, where R is as defined earlier. A halogen group means a
15   fluoro, chloro, bromo or iodo group. It is more preferred that X
is Cl.

If X is OSO_2R, then R is preferably $-CH_3$(mesylate), $-$(p-Me)Ph
(tosylate), $-CF_3$(triflate) or $-C_4F_9$(nonaflate)

20   In the present invention it is also further preferred that the
4,5 fused ring is substituted by $-CO_2R$ in the 2 or 3 position if
it is a benzene ring, and the 2 position if it is a pyrrole ring,
e.g.

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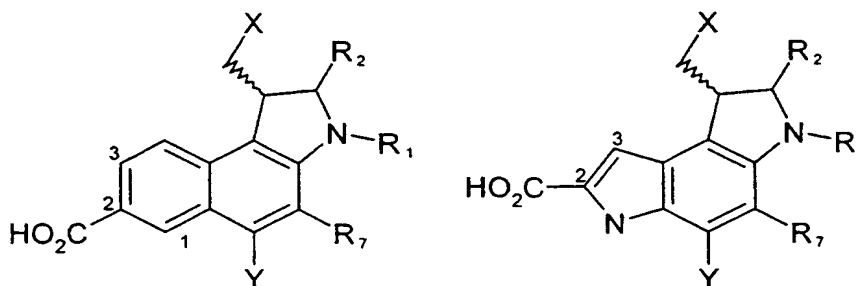
where R_2 and R_7 are preferably H, with the proviso that Y is not
NO_2 when R_1 is Boc, R_2 and R_7 are H, and X is Cl.

30   A third aspect of the present invention relates to compounds of
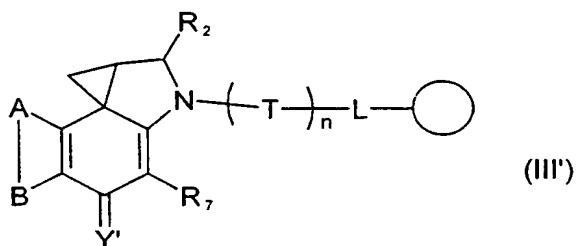the formula **III**:

(III)
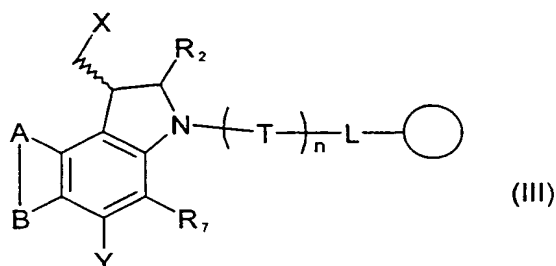
wherein X, Y, A, B, R_2 and R_7 are as defined in the second aspect;
T is a combinatorial unit;
and n is a positive integer, where if n is greater than 1, each T
may be different;
L is a linking group, or less preferably a single bond;
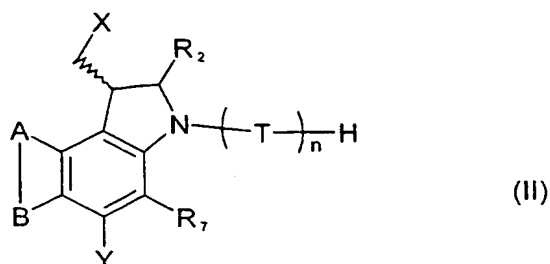and O is a solid support.

A fourth aspect of the present invention relates to compounds of
formula III':

(III')

where A, B, R_2, R_7, T, n, L and O are as defined in the third
aspect; and
Y' is NH, NR, O or S, preferably NH or S.

Compounds of this aspect are obtainable from compounds of the
second aspect by removal of the protecting group preferably
carbamate in Y (if present), or by other appropriate reactions,
e.g. reduction and/or basic conditions. The cyclisation may also
occur spontaneously. A fifth aspect of the present invention
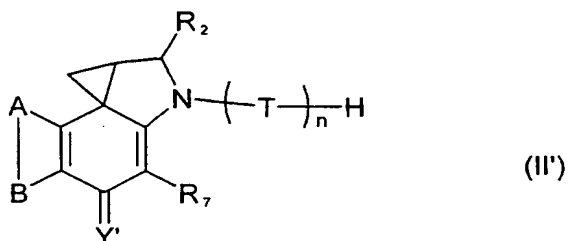relates to compounds of formula II:

(II)

wherein X, Y, A, B, R_2, R_7, T and n are as defined in the third aspect of the invention.

Compounds of this aspect are obtainable by cleavage of the linking group of the appropriate compound of the third aspect.

A sixth aspect of the present invention relates to compounds of the formula **II'**:

(II')

wherein A, B, T, n, R_2 and R_7 are as defined in the fifth aspect of the invention; and
Y' is NH, NR, O, or S, preferably NH or S.

Compounds of this aspect are obtainable by cleavage of the linking group of the appropriate compound of the third aspect (see below).
Alternatively, they are obtainable from compounds of the fourth aspect by removal of the protecting group in Y (if present), or by other appropriate reactions, e.g. reduction and/or basic conditions. The cyclisation may also occur spontaneously.

A seventh aspect of the present invention relates to compounds of formula **V**:

(V)
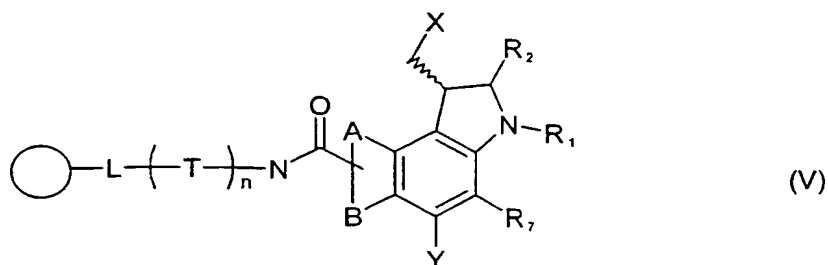
where A, B, Y, R_1, R_2, R_7, and X are defined in the second aspect of the invention, and T, n, L and O are as defined in the third
aspect of the present invention.

An eighth aspect of the present invention relates to compounds of formula **V′**:

(V')

where A, B, R_1, R_2, R_7, T, n, L and O are as defined in the seventh aspect; and
Y′ is NH, NR, O or S, preferably NH or S.

Compounds of this aspect are obtainable from compounds of the seventh aspect by removal of the protecting group in Y (if present), or by other appropriate reactions, e.g. reduction and/or basic conditions. The cyclisation may also occur spontaneously.
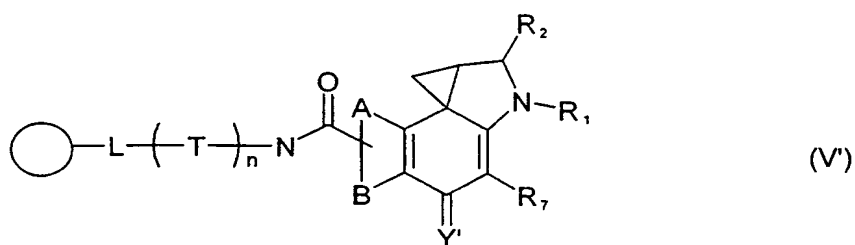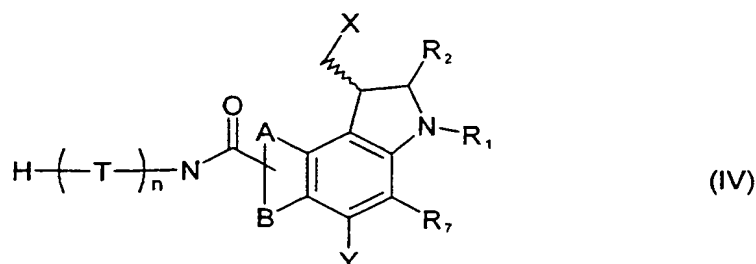
A ninth aspect of the present invention relates to compounds of formula **IV**:
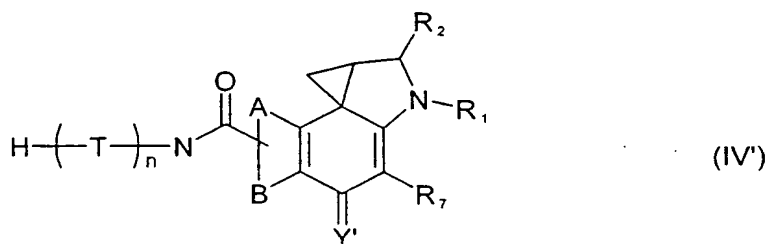

(IV)

wherein A, B, X, Y, T, n, R_1, R_2 and R_7 are as defined in the seventh aspect of the invention.

Compounds of this aspect are obtainable by cleavage of the linking group of the appropriate compound of the seventh aspect.

A tenth aspect of the present invention relates to compounds of formula **IV'** :

(IV')

wherein A, B, T, n, R_1, R_2 and R_7 are as defined in the ninth aspect of the invention; and
Y' is NH, NR, O, or S, preferably NH or S.

Compounds of this aspect are obtainable by cleavage of the linking group of the appropriate compound of the eighth aspect (see below). Alternatively, they are obtainable from compounds of the ninth aspect by removal of the protecting group in Y (if present), or by other appropriate reactions, e.g. by reduction and/or basic conditions. The cyclisation may also occur spontaneously.

An eleventh aspect of the invention is the preparation of compounds according to either the third or seventh aspects by reaction of a compound of formula **VI**:
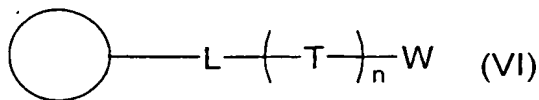
with a compound of formula **I** according to the second aspect, where T, n, L and O are as defined in these aspects, and W is H or an atom or group for providing a functional group capable of reaction with -COOH or $-NH_2$. This reaction will include the necessary protection and deprotection steps so as to selectively achieve a compound according to the third or seventh aspect.

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A twelfth aspect of the invention relates to compounds of formula
VII:

$$\bigcirc - L \left[\left(T' \right)_m T'' \left(T \right)_n \right]_p E \qquad \text{(VII)}$$
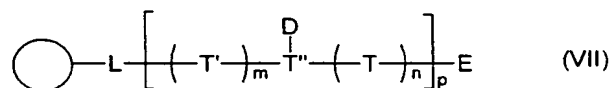
5   wherein:

O, T, and L are as defined in the third aspect of the invention;
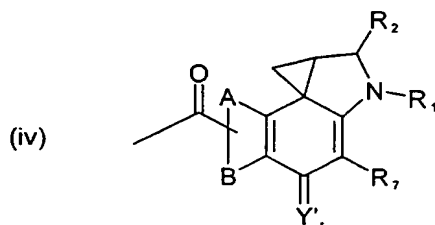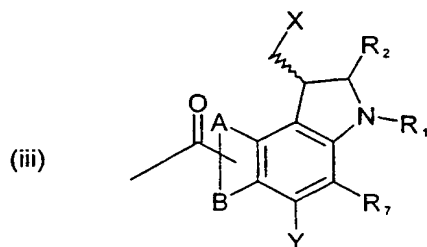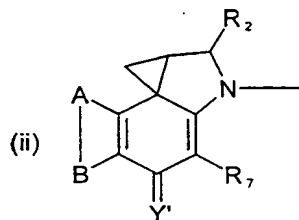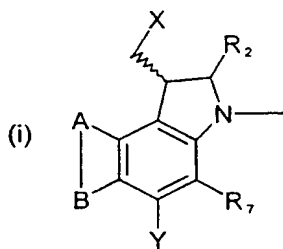
n and m are positive integers, or one of them may be zero;

T' is a combinatorial unit, where each T' may be different if m is
greater than 1;

10   T'' is a combinatorial unit which provides a site for the
attachment of D;

D is selected from:

(a)

(i)

(ii)

(iii)

(iv)

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wherein A, B, Y, Y', R_2 and R_7 are as defined in the second or third aspects of the invention; or

(b)

(i)

(ii)
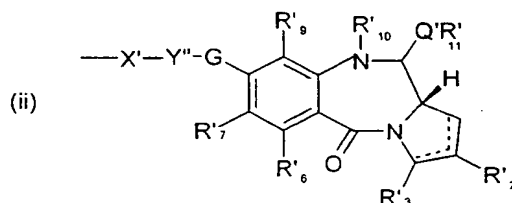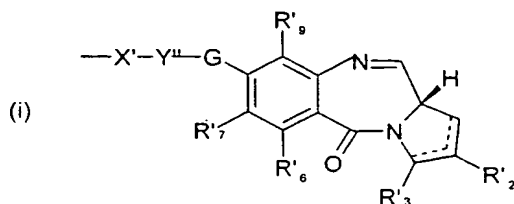
wherein X' is selected from CO, NH, S, or O;

G is O, S, NH, or a single bond;

R'_2 and R'_3 are independently selected from: H, R, OH, OR, =O, =CH-R, $=CH_2$, CH_2-CO_2R, CH_2-CO_2H, CH_2-SO_2R, $O-SO_2R$, CO_2R, COR and CN, and there is optionally a double bond between C_2 and C_3;

R'_6, R'_7, and R'_9 are independently selected from H, R, OH, OR, halo, nitro, amino, Me_3Sn;

R'_{11} is either H or R;

Q' is S, O or NH;

R'_{10} is a nitrogen protecting group;

Y" is a divalent group such that HY = R;

p is a positive integer, where if p is greater than 1, for each repeating unit, the meaning of T, T', T" and D and the values of n and m are independently selected; and

E is selected from the same possibilities as D, provided that at least one group D or E is selected from (a).

If, for example, D terminates with CO then the site on T" may be NH, and if D terminates with NH, S or O, then the site on T" may be CO.

In preferred embodiments of this aspect, one of D and E is selected from (a) and the other is selected from (b).

A thirteenth aspect of the invention relates to compounds of formula (**VIII**):

$$H \left[\left(T' \right)_m \overset{\overset{D}{|}}{T''} \left(T \right)_n \right]_p E \qquad \text{(VIII)}$$
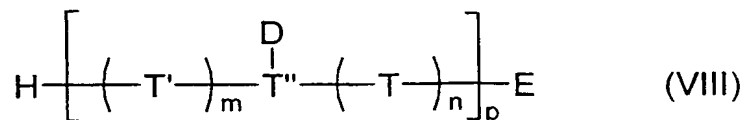
5   wherein L, T, T', T", D, E, n, m and p are as defined in the twelfth aspect of the invention.

In preferred embodiments of this aspect, one of D and E is selected from (a) and the other is selected from (b).

10

A fourteenth aspect of the invention relates to compounds of formula (**IX**):

$$O - L \left[\left(T' \right)_m \overset{\overset{H}{|}}{\underset{q}{\overset{|}{T'''}}} T'' \left(T \right)_n \right]_p E_a \qquad \text{(IX)}$$
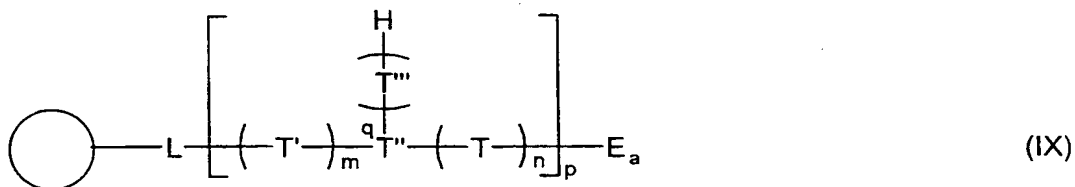
15   wherein O, L, T, T', T", n, m and p are as defined in the twelfth aspect of the invention;

T'" is a combinatorial unit;

q is a positive integer, where if q is greater than 1, each T'"

20 · may be different; and

E_a is selected from the group (a) of E as defined in the twelfth aspect of the invention;

where if p is greater than 1, for each repeating unit the meaning of T, T', T", T"' and the values of n, m and q are independently

25   selected.

A fifteenth aspect of the invention relates to compounds of formula (X):

$$H \underbrace{\left[\left(T' \right)_{m} \underset{\overset{|}{\underset{q}{T'''}}}{\overset{H}{|}} T'' \left(T \right)_{n} \right]_{p}}_{} E_{a} \qquad (X)$$
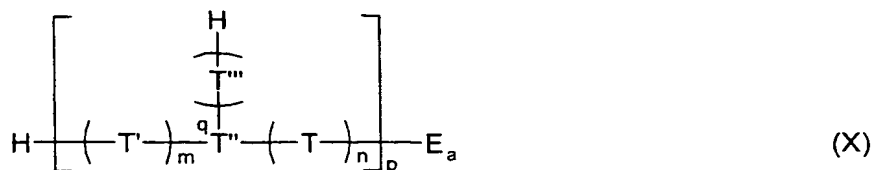
wherein L, T, T', T'', T''', E$_a$, n, m, p and q are as defined in the
fourteenth aspect of the invention.

It is recognised that the compounds of formulae **I, II, II', III,
III', IV, IV', V, V', VII, VIII, IX** and **X** may exist in different
enantiomeric or diastereomeric forms. In such cases it is to be
understood that the above formulae represent any possible
enantiomeric or diastereomeric form or a mixture thereof.

Solid support

The term 'solid support' refers to a material having a rigid or
semi-rigid surface which contains or can be derivatized to contain
reactive functionalities which can serve for covalently linking a
compound to the surface thereof. Such materials are well known in
the art and include, by way of example, silicon dioxide supports
containing reactive Si-OH groups, polyacrylamide supports,
polystyrene supports, polyethyleneglycol supports, and the like.
Such supports will preferably take the form of small beads,
pins/crowns, laminar surfaces, pellets, disks. Other conventional
forms may also be used.

Linker group

The linking groups suitable for the present application are ones
which usually provide in the structure:

$$\bigcirc \!\!-\! L \!-\!$$
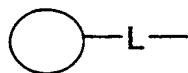
at least one covalent bond which can be readily broken by specific
chemical reactions, such as oxidation (e.g. using DDQ or CAN),
nucleophilic attack (e.g. with an amine on an oxime linker, or an

alkylated sulfamyl linker (or by light, e.g. 365 nm, or changes in pH, e.g. by adding TFA) thereby providing for liberation of compounds free from the solid support. The methods employed to break the covalent bond are selected so as to be specific for the
5 desired bond breakage thereby preventing unintended reactions from occurring elsewhere on the complex. The linking group is selected relative to the synthesis of the compounds to be formed on the solid support so as to prevent premature cleavage of this compound from the solid support as well as to limit interference by any of
10 the procedures employed during compound synthesis on the support.

Examples of resins incorporating cleavable linking groups are set out in the table below, which also indicates the groups that can be immobilised thereon, along with the suggested cleavage methods
15 for the linking group. Such resins are commercially available (e.g. from NovaBiochem).

Linker/Resin Type	Immobilises	Cleavage Method
2-Chlorotrityl chloride	RNH_2, RCO_2H, ROH, RSH	1-50% TFA
Trityl chloride	RNH_2, RCO_2H, ROH, RSH	1-5% TFA
2-Methoxytrityl chloride	RNH_2, RCO_2H, ROH, RSH	1-5%
Rink amide resin	RCO_2H	95% TFA
Sieber amide resin	RCO_2H	1% TFA
4-Sulfamyl- benzoyl	RCO_2H	Alkylation /amines
Wang resin	ROH, ArOH, RNH_2, RCO_2H	15-95% TFA or DDQ or CAN
HMPB-BHA	ROH, ArOH, RCO_2H	1% TFA
Bromoethyl photolinker	RNH_2, RCO_2H, ROH, RSH	hv
Hydroxy ethyl photolinker	RCO_2H	hv
Aminoethyl photolinker	RCO_2H	hv

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Structures

[RESIN] — structure — Cl, D
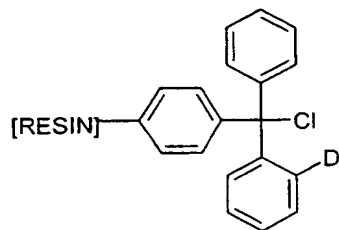
D = Cl: 2-chlorotrityl chloride type
D = H: trityl chloride type
D = OMe: 2-methoxytrityl chloride

OMe NHFMOC

MeO   O—[RESIN]   Rink amide type

NHFMOC

O—[RESIN]   Sieber amide type

H_2N—S—structure—$\overset{O}{\underset{O}{S}}$ ... $\overset{O}{\parallel}$ ...—[RESIN]   4-sulfamyl-benzoyl type

HO—structure—O—[RESIN]   Wang type

MeO
HO—structure—O—structure—[RESIN]   HMPB-BHA type
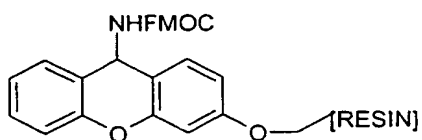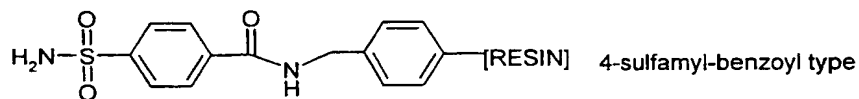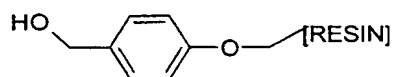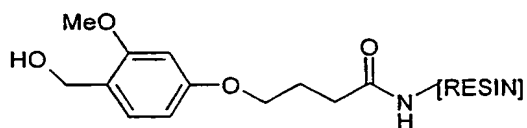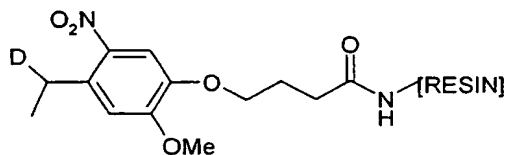
O_2N
D—structure—OMe—O—structure—[RESIN]   D = NH_2:amino-ethyl type
D = OH: hydroxy-ethyl type
D = Br: bromo ethyl type

For CPI precursors the most preferred linking group is one which may be deemed photolytically cleavable. Further, the Rink amide linker is particularly suitable.

It is also possible that the linking group is a simple functionality provided on the solid support, e.g. amine, and in this case the linking group may be not be readily cleavable. This type of linking group is useful in the synthesis of large split and mix libraries which will be subjected to on-bead screening (see below), where cleavage is unnecessary. Such resins are commercially available from a large number of companies including NovaBiochem, Advanced ChemTech and Rapp Polymere. These resins include amino-Tentagel and aminomethylated polystyrene resin.

Combinatorial Unit

The term 'combinatorial unit' means any monomer unit which can be used to build a chain attached to the solid support, usually by a linking group. Usually combinatorial units will comprise at least two different functional groups to provide them with this chain building ability. For example, amino acids comprise both carboxylic acid and amine moieties. Sometimes the combinatorial unit may require more than two functionalities, e.g. if it has to bond to a further moiety. Examples of molecules suitable for such chain building are found in Schreiber et al. (*JACS*, 120. 1998, pp.23-29), which is incorporated herein by reference. An important example of a unit is an amino acid residue. Chains may be synthesised by means of amine-protected amino acids. Fmoc protected amino-acids are available from a number of sources, such as Sigma and Nova Biochem. Both natural and unnatural amino acids can be used, e.g. D- and L-amino acids and heterocyclic amino acids. In particular, heterocyclic amino acids of the type found in the construction of netropsin and distamycin are of interest because of their DNA-recognition properties.

Amine units can be used to make up peptoids: see Soth, M.J. and Nowick, J.S. 1997, Unnatural oligomer libraries, *Curr. Opin, Chem.*

Biol. 1, no. 1: 120-129; Zuckermann *et al.*, 1994, Discovery of Nanomolecular Ligands for 7-Transmembrane G-Protein-Coupled Receptors from a Diverse N-(Substituted)glycine Peptoid Library, *Journal of Medicinal Chemistry* 37: 2678-85; Figliozzi, GMR *et al.*, 5   1996, Synthesis of N-substituted Glycine Peptoid Libraries, *Methods in Enzymology*, 267: 437-47; Simon, R J *et al.*, 1992, Peptoids: A Modular Approach to Drug Discovery, *Proc. Natl. Acad. Sci. USA*, 89:9367-71; which are all incorporated herein by reference.

10

Other combinatorial units include PNAs (peptidonucleic acids): PE Nielsen, *et al*, *Science*, 1991, 254, 1497; M Egholm, *et al*, *Nature*, 1993, 365, 566; M Egholm *et al*, *JACS*, 1992, 114, 1895; S C Brown, *et al*, *Science*, 1994, 265, 777; 5. K Saha, *et al*, *JOC*, 1993, 58, 15   7827; oligoureas: Burgess K, *et al*, 1995, Solid Phase Synthesis of Unnatural Biopolymers Containing Repeating Urea Units. *Agnew. Chem. Int. Ed. Engl* 34, no. 8:907; Burgess K, *et al*, 1997, Solid Phase Synthesis of Oligoureas; *Journal of the American Chemical Society* 119: 1556-64; and oligocarbamates: Moran E J *et al*, 1995, 20   Novel Biopolymers for Drug Discovery. *Biopolymers (Peptide Science); John Wiley and Sons* 37: 213-19; Cho C Y *et al*, 1993, An Unnatural Biopolymer. *Science* 261: 1303-5; Paikoff S F *et al*, 1996, The Solid Phase Synthesis of N-Alkylcarbamate Oligomers. *Tetrahedron Letters* 37, no. 32: 5653-56. All of these documents 25   are incorporated herein by reference.

A further aspect of the present invention relates to combinatorial units having formula **I**, where the fused ring collectively represented by A and B is substituted by CO_2R.

30

Further combinatorial units of relevance to this invention are those of formulae (**XIa/b**):

(XIa)

(XIb)

5
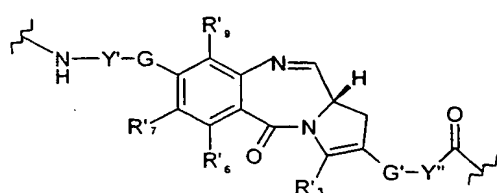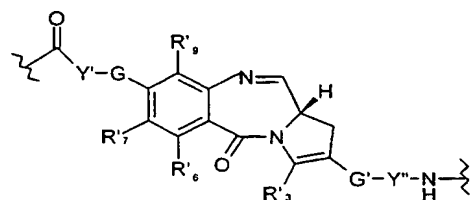
wherein R'_3, R'_6, R'_7, R'_9, G and Y' are as defined in the twelfth aspect of the invention, and G' and Y" are independently selected from the possible groups for G and Y' respectively. In order to synthesise combinatorial chains containing such combinatorial units,

10   the units may need to be joined to the chain in their protected form (see definition of groups D(b) above). It is possible that the combinatorial units may remain in their protected form until the compound is cleaved from the solid support, or until the other components in the compound are deprotected.

15

Combinatorial units may also be of formula I with substituents as defined in the second aspect of the invention, where the fused ring, represented by -A-B-, bears a substituent which is $-CO_2H$ or $-CO_2R$.

20   The present invention relates to libraries, or collections, of compounds, all of which are represented by a single one of the formulae **II**, **II'**, **III**, **III'**, **IV**, **IV'**, **V**, **V'**, **VII**, **VIII**, **IX** or **X**. The diversity of the compounds in a library may reflect the presence of compounds differing in the identities of one or more

25   of (as appropriate) X, Y, Y', A, B, R_2, R_7, X', Y", G, R'_2, R'_3, R'_6, R'_7, R'_9, R'_{10}, Q', R'_{11} and/or in the identities of the combinatorial units T, T', T" and T'" (when present). The number of members in the library depends on the number of variants, and the number of possibilities for each variant. For example, if it

30   is the combinatorial units which are varied, and there are 3 combinatorial units, with 3 possibilities for each unit, the library will have 27 compounds. 4 combinatorial units and 5 possibilities for each unit gives a library of 625 compounds. If,

for instance, there is a chain of 5 combinatorial units with 17
possibilities for each unit, the total number of members in the
library would be 1.4 million. A library may therefore comprise
more than 1 000, 5 000, 10 000, 100 000 or a million compounds,

5   which may be arranged as described below.
       In the case of free compounds (formulae **II, II′, IV, IV′, VIII, X**)
       the individual compounds are preferably in discrete volumes of
       solvents, e.g. in tubes or wells. In the case of bound compounds
       (formulae **III, III′, V, V′, VII, IX**) the individual compounds are

10   preferably bound at discrete locations, e.g. on respective
       pins/crowns or beads. The library of compounds may be provided on
       a plate which is of a suitable size for the library, or may be on
       a number of plates of a standard size, e.g. 96 well plates. If
       the number of members of the library is large, it is preferable

15   that each well on a plate contains a number of related compounds
       from the library, e.g. from 10 to 100. One possibility for this
       type of grouping of compounds is where only a subset of the
       combinatorial units, or substituents, are known and the remainder
       are randomised; this arrangement is useful in iterative screening

20   processes (see below). The library may be presented in other
       forms that are well-known.

       A further aspect of the present invention is a method of preparing
       a diverse collection, or library of compounds, as discussed above.

25   If the diversity of the library is in the combinatorial units,
       then the library may be synthesised by the stepwise addition of
       protected combinatorial units to a CPI/CBI precursor core, each
       step being interposed by a deprotection step. Such a method is
       exemplified later. Libraries of this type can be prepared by the

30   method known as "split and mix" which is described in Furka, A;
       Sebestyen, F; Asgedom, M and Dibo, G; General Method of Rapid
       Synthesis of Multicomponent Peptide Mixtures; International
       Journal of Peptide and Protein Research; 1991, **37**, 487-193, which
       is incorporated herein by reference. If the diversity of the

35   library is in the substituent groups, the library may be
       synthesised by carrying out the same synthetic methods on a
       variety of starting materials or key intermediates, which already
       possess the necessary substituent patterns.

22

The present invention also relates to a method of screening the
compounds of formula **II, II′, III, III′, IV, IV′, V, V′, VII,
VIII, IX** or **X** to discover biologically active compounds. The

5   screening can be used to assess the binding interaction with
nucleic acids, e.g. DNA or RNA, or proteins, or to assess the
affect of the compounds against protein-protein or nucleic
acid-protein interactions, e.g. transcription factor DP-1 with
E2F-1, or estrogen response element (ERE) with human estrogen

10   receptor (a 66 kd protein which functions as a hormone-activated
transcription factor, the sequence of which is published in the
art and is generally available). The screening may also be used
to assess the cytotoxicity of the compounds against a variety of
cell lines. The screening can be carried out by bringing the

15   target macromolecules into contact with individual compounds or
the arrays or libraries of individual compounds described above,
and selecting those compounds, or wells with mixtures of
compounds, which show the strongest effect.

20   This effect may simply be the cytotoxicity of the compounds in
question against cells or the binding of the compounds to nucleic
acids. In the case of protein-protein or nucleic acid-protein
interactions, the effect may be the disruption of the interaction
studied.

25

The binding of the compounds to nucleic acids may be assessed by
labelling oligomers which contain a target sequence, and measuring
the amount of labelled oligomers that bind to the compounds
tested. The labelling may either be radiolabelling, or

30   alternatively be labels detectable under visible or ultra-violet
light. If this latter form of screening is carried out on
compounds bound to solid supports which are in separate locations,
the screening for results can be carried out visually under a
microscope. A similar technique is described in detail in

35   DNA-Binding ligands from peptide libraries containing unnatural
amino acids, Lescrinier *et al.*, *Chem Eur J*, 1998, 425-433. These
techniques are particularly suited to a one-step screening of a

complete library of compounds, especially a large library made by the "split and mix" method described above.

Protein-protein interactions can be measured in a number of ways,
5   e.g. FRET (fluorescence resonance energy transfer) which involves labelling one of the proteins with a fluorescent donor moiety and the other with an acceptor which is capable of absorbing the emission from the donor; the fluorescence signal of the donor will be altered depending on the interaction between the two proteins.
10 Another method of measuring protein-protein interactions is by enzymatic labelling, using, for example, horseradish peroxidase.

The screening process may undergo several iterations by selecting the most active compounds, or group of compounds, tested in each
15 iteration; this is particular useful when testing arrays of wells which include mixtures of related compounds. Furthermore, if the wells contain compounds for which only a subset of the combinatorial units, or substituents, are known, but the rest are randomised, subsequent iterations can be carried out by
20 synthesising compounds possessing the selected known (and successful) combinatorial unit, or substituent, pattern, but with further specified combinatorial units, or substituents, replacing the previously randomised combinatorial units, or substituents, adjacent the already known pattern; the remaining combinatorial
25 units, or substituents, are randomised as in the previous iteration. This iterative method enables the identification of active members of large libraries without the need to isolate every member of the library.

30 A further feature of this aspect is formulation of selected compound or compounds with pharmaceutically acceptable carriers or diluents.

In yet further aspects, the invention provides a pharmaceutical
35 composition comprising a compound of formula II, II', IV, IV', VIII or X and a pharmaceutically acceptable carrier or diluent; and the use of a compound of formula II, II', IV, IV', VIII or X in the manufacture of a medicament for the treatment of a

gene-based disease, or a bacterial, parasitic or viral infection.
Gene-based disease include neoplastic disease and, for example,
Alzheimer's disease.

5   Compounds of formula **II**, **II'**, **IV**, **IV'**, **VIII** or **X** may be used in a
method of therapy against a gene-based disease, such as cancer or
Alzheimer's disease, or a viral, parasitic or bacterial infection.

     Another aspect of the present invention relates to the use of
10   compounds of formula **III**, **III'**, **V**, **V'**, **VII** or **IX** in diagnostic
methods. A compound of formula **III**, **III'**, **V**, **V'**, **VII** or **IX** which
binds to an identified sequence of DNA or a protein known to be an
indicator of a medical condition can be used in a method of
diagnosis. The method may involve passing a sample, e.g. of
15   appropriately treated blood or tissue extract, over an immobilised
compound of formula **III**, **III'**, **V**, **V'**, **VII** or **IX**, for example in a
column, and subsequently determining whether any binding of target
DNA to the compound of formula **III**, **III'**, **V**, **V'**, **VII** or **IX** has
taken place. Such a determination could be carried out by passing
20   a known amount of labelled target DNA known to bind to compound
III, **III'**, **V**, **V'**, **VII** or **IX** through the column, and calculating
the amount of compound **III**, **III'**, **V**, **V'**, **VII** or **IX** that has
remained unbound.

25   A further aspect of the present invention relates to the use of
compounds of formula **II**, **II'**, **IV**, **IV'**, **VIII** or **X** in target
validation. Target validation is the disruption of an identified
DNA sequence to ascertain the function of the sequence, and a
compound of formula **II**, **II'**, **IV**, **IV'**, **VIII** or **X** can be used to
30   selectively bind an identified sequence, and thus disrupt its
function.

     Another aspect of the present invention relates to the use of
compounds of formulae **II**, **II'**, **IV**, **IV'**, **VIII** or **X** in functional
35   genomics to ascertain the biological function of genes, by
blocking this biological action.

25

Preferred Synthetic Strategies

Compounds of formula **I** can be synthesised by applying the methods
described below. A review of methods of synthesising CPIs was

5   carried out by Boger (Boger, D.L. et al., J.A. *Chem. Rev.* **1997,**
97, 787-828)

Boger Synthesis of *N*-BOC-CBI

Selective C4 iodination of *N*-BOC-4-(benzyloxy)naphthylamine,

10   readily accessible in three steps from the commercially available
1,3-dihydroxynaphthalene, followed by N-alkylation with allyl
bromide provided the required substrate for the 5-*exo-trig* aryl
radical-alkene cyclization (Boger, D.L. et al., O. *J. Am. Chem.
Soc.* **1989,** *111,* 6461-6463. Boger, D.L. at al., O. *J. Org. Chem.*

15   **1990,** *55,* 5823-5832)(Scheme 1). Treatment with Bu_3SnH–TEMPO
(Boger, D.L.; McKie, J.A. *J. Org. Chem.* **1995,** *3,* 1429-1453) and
subsequent reduction with Zn afforded the 3-
(hydroxymethyl)indoline CBI precursor. Conversion to the primary
chloride and catalytic hydrogenolysis of the benzyl ether,

20   followed by direct resolution on a semipreparative chiral HPLC
column afforded both enantiomers (Boger, D.L.; Yun, W. *J. Am.
Chem. Soc.* **1994,** *116,* 7996-8006). Subsequent spirocyclization
completed the synthesis.

Scheme 1
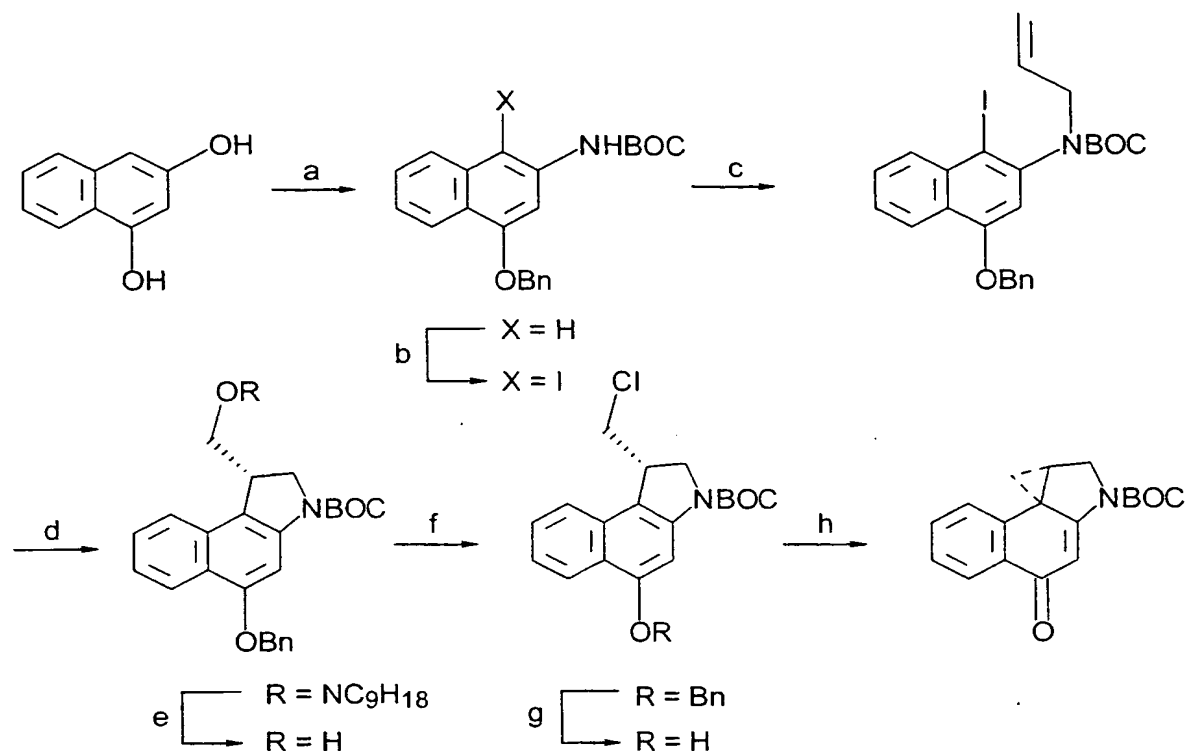
Reagents and conditions: **a**: NH_3, BOC_2O; BnBr; **b**: NIS; **c**: AllylBr, NaH; **d**: Bu_3SnH, TEMPO; **e**: Zn; **f**: Ph_3P-CCl_4; **g**: H_2, Pd-C; **h**: NaH

Cava synthesis of CBI

Cava's route to CBI uses a heterocyclization procedure involving a heterostilbene to establish the tricyclic CBI core, followed by introduction of an additional functionalized carbon necessary for formation of the cyclopropane (Drost, K.J.; Cava, M.P. *J. Org. Chem.* **1991**, *56*, 2240-2244)(Scheme 2).
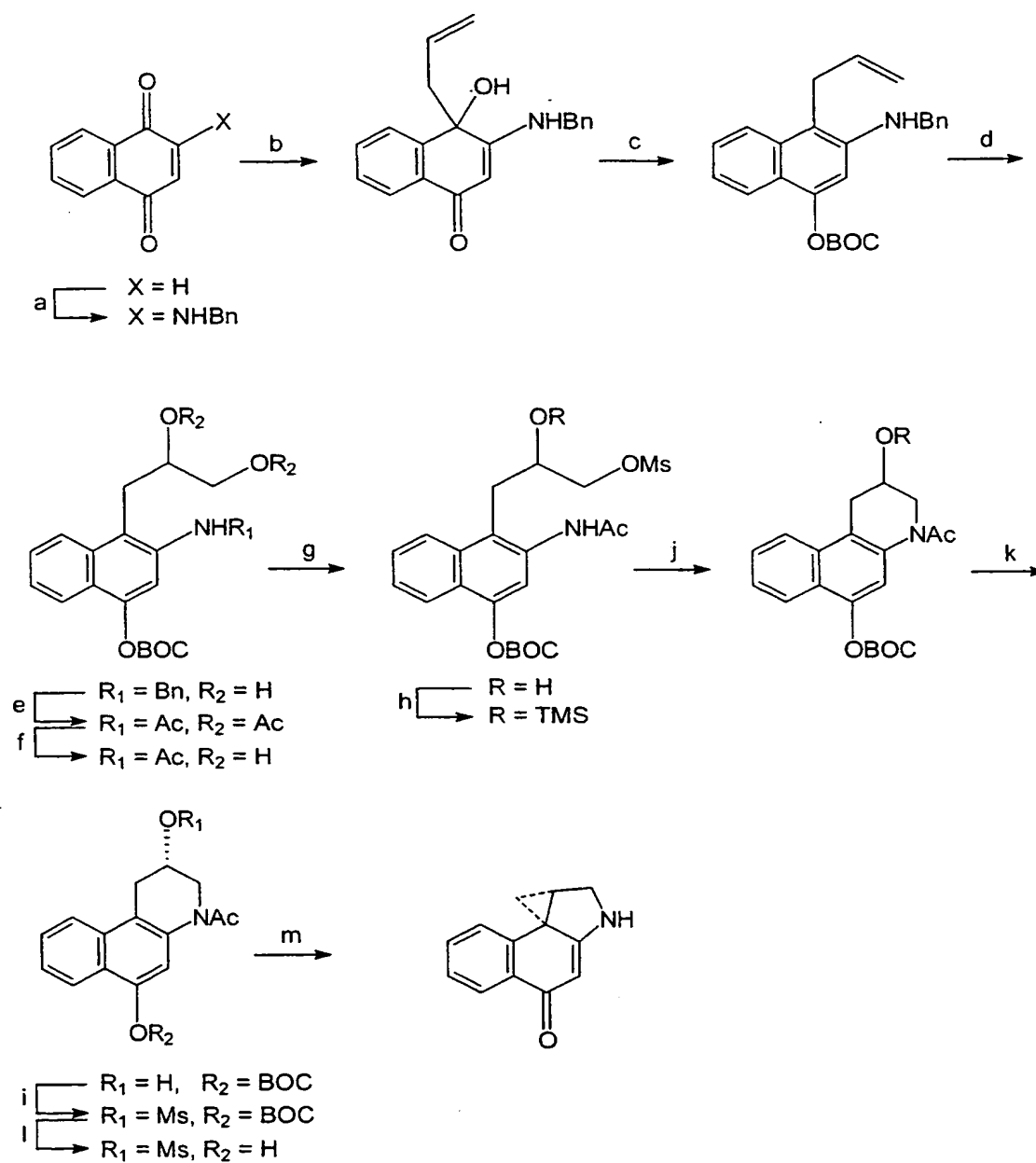
27

Scheme 2
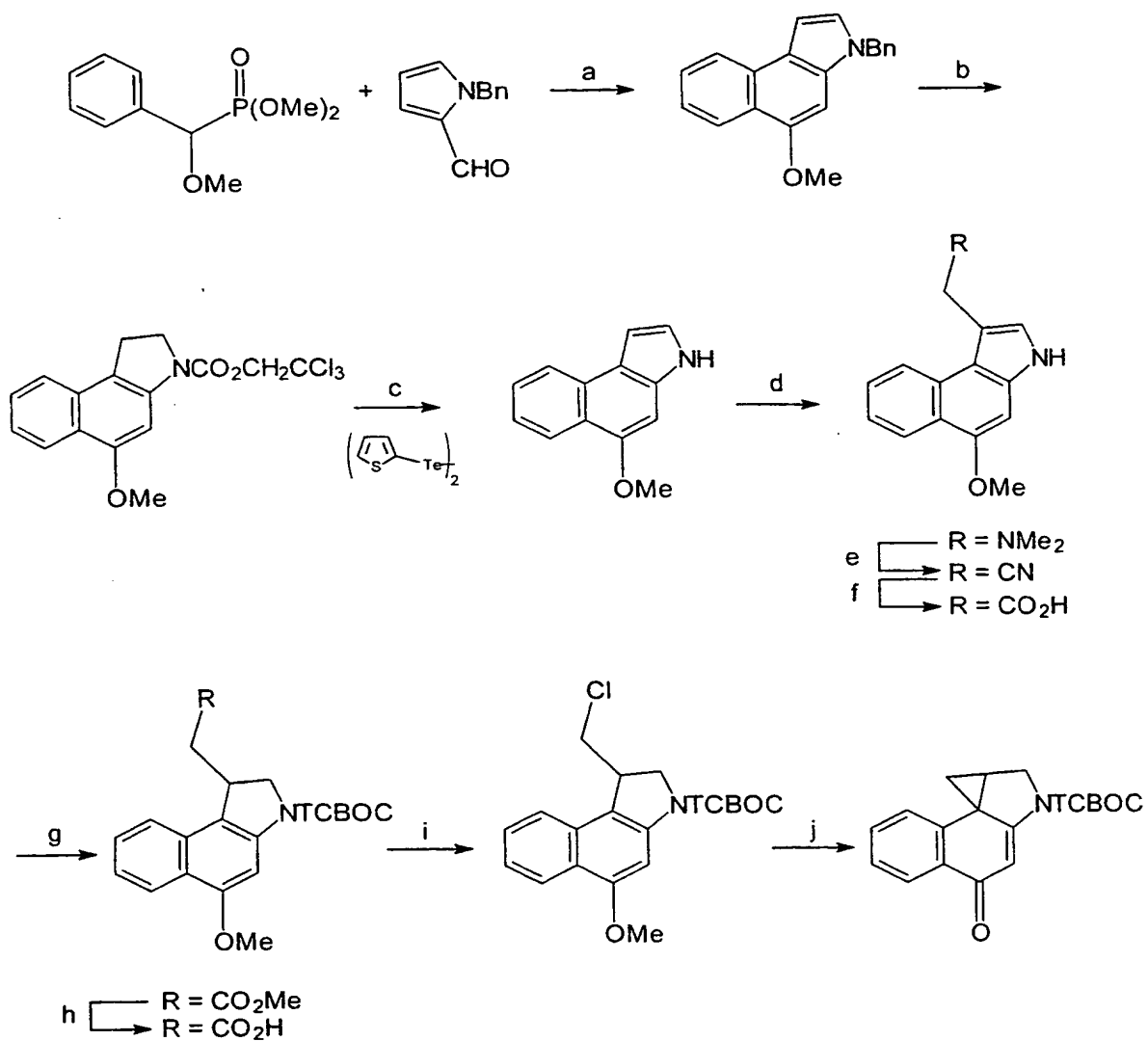
Reagents and conditions: a: 1) t-BuOK; 2) p-nitrobenzoic acid, Et_3N, Pd-C, hn; b: 1) $NaBH_3CN$; 2) $ClCO_2CH_2CCl_3$; c: 1) DDQ; 2) $NaBH_4$; d: Me_2NH, CH_2O; e: CH_3I; NaCN; f: NaOH; g: 1) MeO_2CCl, Et_3N; 2) $NaBH_3CN$; 3) TCBOC-Cl, Et_3N; h: NaOH; i: 1) $(COCl)_2$; 2) mercaptopyridine-N-oxide, DMAP, CCl_4; j: $BCl_3 \cdot SMe_2$; Et_3N

Thus, photolysis of the alkene formed from condensation of the α-methoxybenzyl phosphonate with N-benzylpyrrole-2-carboxaldehyde in the presence of Pd-C provided the tricylic CBI core (Rawal, V.H.; Jones, R.J.; Cava, M.P. *J. Org. Chem.* **1987**, *52*, 19-28). A four step debenzylation sequence followed by a regioselective Mannich alkylation was employed to provide the key CBI intermediate.

Aristoff synthesis of CBI (Scheme 3)

An alternative approach to the CBI subunit was described by Aristoff and coworkers (Aristoff, P.A.; Johnson, P.D.; Sun, D. *J. Med. Chem.* **1993**, *36*, 1956-1963. Aristoff, P.A.; Johnson, P.D. *J. Org. Chem.* **1992**, *57*, 6234-6239). 1-Allyl-2-(benzylamino)-1-hydroxydihydronaphthalenone was prepared in two steps from 1,4-naphthoquinone. Reduction and re-aromatization was accomplished by treatment with BOC_2O followed by sodium dithionite. The racemic diol was prepared by OsO_4-catalyzed dihydroxylation. Deprotection of the benzylamine, N- and O-acetylation, acetate hydrolysis, followed by selective mesylation of the primary alcohol and TMS ether protection of the secondary alcohol preceded 6-membered ring closure upon treatment with NaH. Alcohol deprotection and resolution by chromatographic separation of the diastereomeric (R)-O-acetylmandelate esters provided the optically active materials.

Scheme 3

a ⌐── X = H
  └─➤ X = NHBn

e ── R₁ = Bn, R₂ = H
f ⌐─➤ R₁ = Ac, R₂ = Ac
  └─➤ R₁ = Ac, R₂ = H

h ⌐ R = H
  └─➤ R = TMS

i ⌐── R₁ = H, R₂ = BOC
l ⌐─➤ R₁ = Ms, R₂ = BOC
  └─➤ R₁ = Ms, R₂ = H

Reagents and conditions: **a**: $BnNH_2$; **b**: Allyl MgBr; **c**: BOC_2O; $Na_2S_2O_4$; **d**: OsO_4, NMO; **e**: HCO_2H, Pd-C; Ac_2O; **f**: K_2CO_3; **g**: TMSCl, pyridine; **h**: NaH; **j**: K_2CO_3; **k**: (R)-O-acetylmandelic acid, EDCI; Resolution; K_2CO_3; **i**: MsCl, Et_3N; **l**: TFA; **m**: NaH

Primary alcohol activation, BOC deprotection, and transannular cyclization upon treatment with NaH provided the CBI accompanying hydrolysis of the intermediate N-Ac-CBI by water present in the reaction mixture.

Any of these three routes may be adapted to synthesise compounds according to the first aspect of the present invention, for example by starting with appropriately substituted starting materials (which may be protected), or by introducing substituents at a later stage.

Preferred Synthetic Strategies of
pyrrolo[2,1-c][1,4]benzodiazepines (also see WO 00/12506)
A key step in a preferred route to compounds corresponding to the group D(b) of the eleventh aspect of the invention or combinatorial units of formulae **XIa** and **XIb** is a cyclisation process to produce the B-ring, involving generation of an aldehyde (or functional equivalent thereof) at what will be the 11-position, and attack thereon by the pro-10-nitrogen:
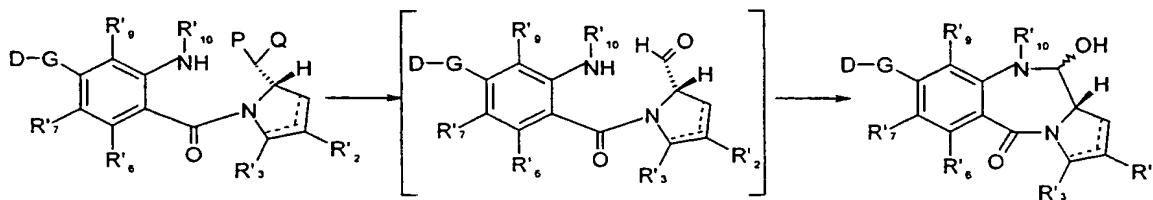
In this structure, D represents XY, or a masked form thereof. The "masked aldehyde" -CPQ may be an acetal or thioacetal (possibly cyclic), in which case the cyclisation involves unmasking. Alternatively, the masked aldehyde may be an aldehyde precursor

such as an alcohol -CHOH, in which case the reaction involves oxidation, e.g. by means of TPAP or DMSO (Swern oxidation).

The masked aldehyde compound can be produced by condensing a corresponding 2-substituted pyrrolidine with a 2-nitrobenzoic acid:
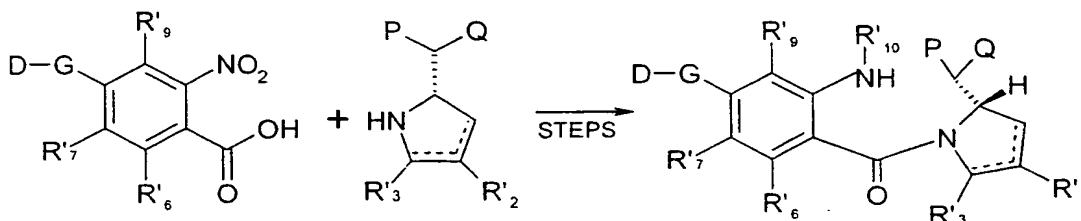
The nitro group can then be reduced to $-NH_2$ and protected by reaction with a suitable agent, e.g. a chloroformate, which provides the removable nitrogen protecting group in the desired compound.

A process involving the oxidation-cyclization procedure is illustrated in scheme 4 (an alternative type of cyclisation will be described later with reference to scheme 5).
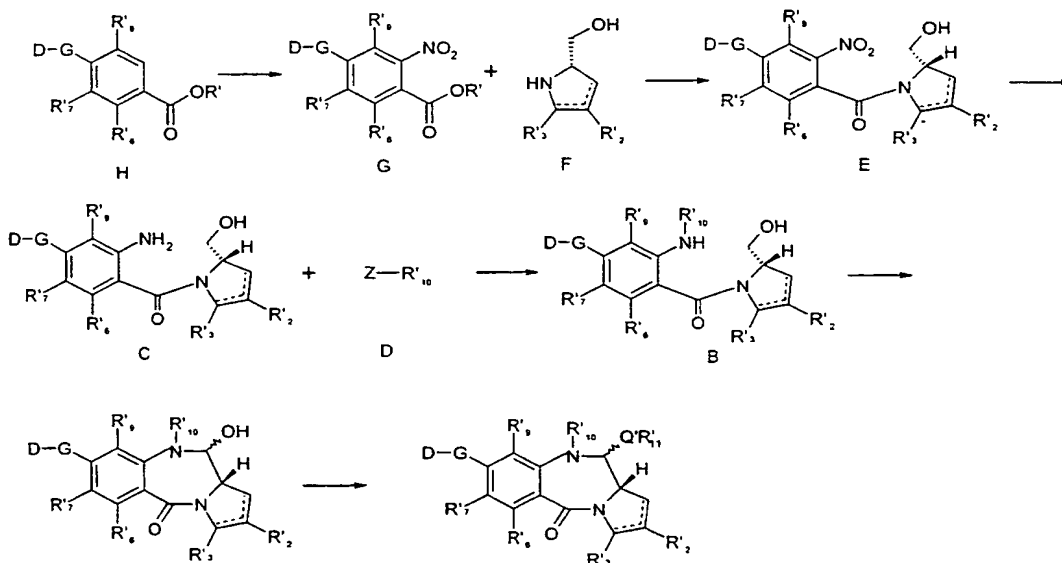
Scheme 4

If R'_{11} is other than hydrogen, the final compound may be prepared
by direct etherification of the alcohol. Compounds with $Q' = S$
can be prepared by treatment of the corresponding alcohol with
5      $R'_{11}SH$, and a catalyst (usually a Lewis Acid such as Al_2O_3). If Q'
= NH, then these compounds can be prepared by reacting the alcohol
with an amine $R'_{11}NH$ and a catalyst (usually a Lewis Acid).

Exposure of the alcohol **B** (in which the 10-nitrogen is generally
10   protected as an amide carbamate) to tetrapropylammonium
perruthenate (TPAP)/N-methylmorpholine N-oxide (NMO) over A4
sieves results in oxidation accompanied by spontaneous B-ring
closure to afford the desired product. The TPAP/NMO oxidation
procedure is found to be particularly convenient for small scale
15   reactions while the use of DMSO-based oxidation methods,
particularly Swern oxidation, proves superior for larger scale
work (e.g. > 1 g).

The uncyclized alcohol **B** may be prepared by the addition of a
20   nitrogen protecting reagent of formula **D**, which is preferably a
chloroformate or acid chloride, to a solution of the amino alcohol
C, generally in solution, generally in the presence of a base such
as pyridine (preferably 2 equivalents) at a moderate temperature
(e.g. at 0°C). Under these conditions little or no O-acylation is
25   usually observed.

The key amino alcohol **C** may be prepared by reduction of the
corresponding nitro compound **E**, by choosing a method which will
leave the rest of the molecule intact. Treatment of **E** with tin
30   (II) chloride in a suitable solvent, e.g. refluxing methanol,
generally affords, after the removal of the tin salts, the desired
product in high yield.

Exposure of **E** to hydrazine/Raney nickel avoids the production of
35   tin salts and may result in a higher yield of **C**, although this
method is less compatible with the range of possible C and A-ring
substituents. For instance, if there is C-ring unsaturation

(either in the ring itself, or in R_2 or R_3), this technique may be unsuitable.

The nitro compound of formula **E** may be prepared by coupling the appropriate o-nitrobenzoyl chloride to a compound of formula **F**, e.g. in the presence of K_2CO_3 at -25°C under a N_2 atmosphere. Compounds of formula **F** can be readily prepared, for example by olefination of the ketone derived from L-trans-4-hydroxy proline. The ketone intermediate can also be exploited by conversion to the enol triflate for use in palladium mediated coupling reactions.

The o-nitrobenzoyl chloride is synthesised from the o-nitrobenzoic acid (or alkyl ester after hydrolysis) of formula **G**, which itself is prepared from the vanillic acid (or alkyl ester) derivative **H**. Many of these are commercially available and some are disclosed in Althuis, T. H. and Hess, H. J., *J. Medicinal Chem*, 20(1), 146-266 (1977).

Alternative Cyclisation (Scheme 5)
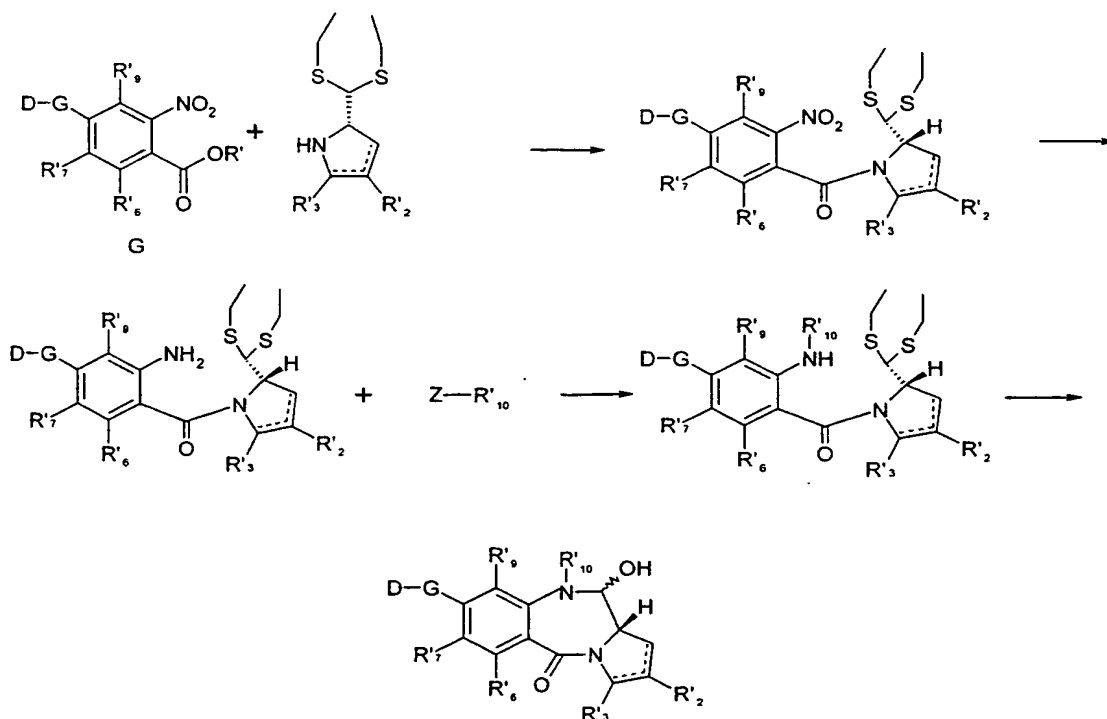
Scheme 5

In scheme 4, the final or penultimate step was an oxidative
cyclisation. An alternative approach, using thioacetal
coupling/unmasking, is shown in scheme 5. Mercury-mediated
5   unmasking causes cyclisation to the desired compound.

The thioacetal intermediates may be prepared as shown in scheme 2:
the thioacetal protected C-ring [prepared via a literature method:
Langley, D.R. & Thurston, D.E., *J. Organic Chemistry*, **52**, 91-97
10   (1987)] is coupled to the *o*-nitrobenzoic acid (or alkyl ester
after hydrolysis) **G** using a literature procedure. The resulting
nitro compound cannot be reduced by hydrogenation, because of the
thioacetal group, so the tin(II) chloride method is used to afford
the amine. This is then N-protected, e.g., by reaction with a
15   chloroformate or acid chloride, such as *p*-
nitrobenzylchloroformate.

Acetal-containing C-rings can be used as an alternative in this
type of route with deprotection involving other methods including
20   the use of acidic, or perhaps Lewis Acid, conditions.

In the above synthesis schemes, the derivatisation of the A-ring
is shown as being complete before the compounds are attached to
the solid support. This is preferred is the substituents are
25   groups such as alkoxy or nitro. On the other hand, substituent
groups such as alkyl or alkenyl could be added to the A-ring after
the coupling of the compound to the solid support. This may be
achieved by R'_6, R'_7, or R'_9 being easily replaceable groups, such
as a halogen atom.
30

An alternative synthesis approach to those detailed above is to
protect the Pro N10 position on the component which will form the
A-ring before joining the component which will form the C-ring.

35   Embodiments of the present invention will now be described by way
of example.

Example 1:   CBI bound to TG-Carboxy resin

Activation of TG-Carboxy resin with 3-hydroxy-1,2,3-benzotriazin-4(3H)-one **(HO-Dhbt)**
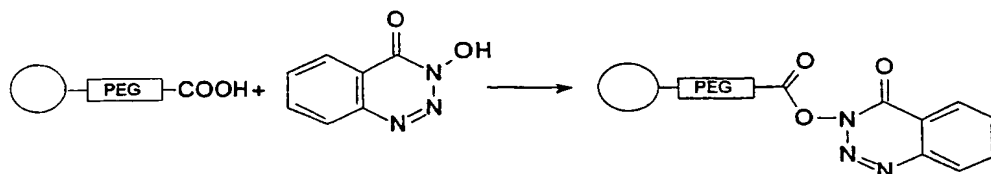

TG Carboxy resin (Nova-Biochem, 80 mg 0.020 mmol) was suspended and swollen in DMF (1 mL) for 10 minutes.   DMF was removed by suction and the resin treated with diisopropyl carbodiimide (DIC) (34 mmL, 10 eq)in of DMF (1 mL).   The mixture was shaken for 15 minutes at -15°C (acetone-ice bath), then rinsed with DMF (2 x 1 mL).

The resin was suspended in DMF(1 mL), cooled to -10°C and treated with HO-Dhbt(33 mg, 10 eq).   The mixture was shaken for 30 minutes at -10°C, followed by 4 hours shaking at 0°C and was then allowed to stand at 0°C overnight.

The reaction mixture was filtered and the resin washed with DMF (2 x 1 mL), dichloromethane (2 x 1 mL), MeOH (2 x 1 mL) and diethyl ether (1 mL).   The activated resin was stored in freezer until ready for use.

Coupling of O-Dhbt activated TG-Carboxy resin with *seco*-CBI, in presence of DIPEA
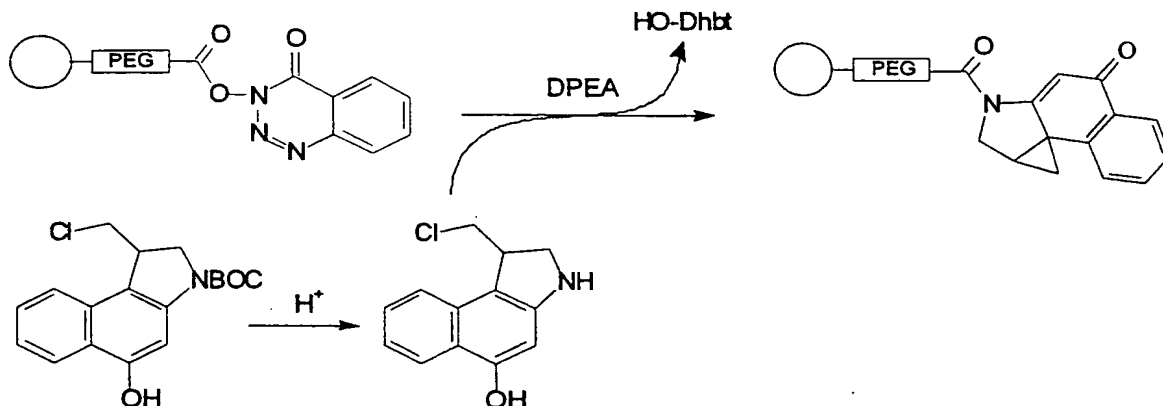

5   Seco-CBI has a very limited storage time, and for this reason BOC-*seco*-CBI is deprotected immediately before coupling to the activated resin. BOC-*seco*-CBI (20 mg 0.06 mmol) was dissolved in HCl in anhydrous ethyl acetate (3M, 2.8 mL). The mixture was kept at 0°C for 30 minutes and then allowed to warm to room temperature
10   over 30 minutes. TLC (4:1 petroleum ether:diethyl ether) revealed complete reaction; the reaction mixture was evaporated under reduced pressure and stored at -20°C.

O-Dhbt activated resin (40 mg) was treated with *seco*-CBI dissolved
15   in DMF (1 mL). DIPEA (3 eq, 7.7 mg, 10 mmL) was added to the suspension and the mixture was shaken for 30 minutes at room temperature. The reaction mixture was filtered and the resin was washed with DMF (1 mL), dichloromethane (1 mL), methanol (1 mL) and diethyl ether (2 x 1 mL). After thorough evaporation of solvents the
20   resin was stored at -20°C.

Example 2: seco-CBI bound to TG-Carboxy resin

Coupling of O-Dhbt activated TG-Carboxy resin with *seco*-CBI, in absence of base
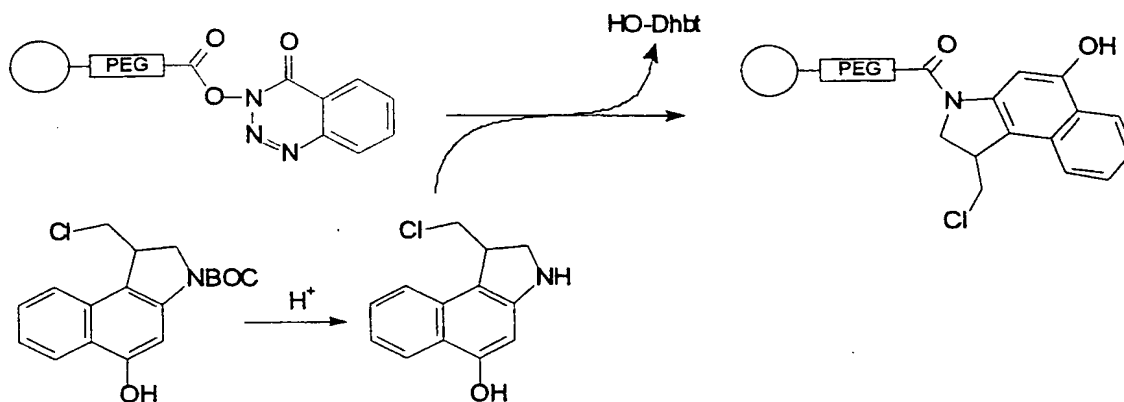
BOC-*seco*-CBI (20 mg, 0.06 mmol) was dissolved in HCl solution in anhydrous ethyl acetate (3M, 2.8 mL). The mixture was kept at 0°C for 30 minutes and then allowed to warm to room temperature over 30 minutes. TLC (4:1 = petroleum ether:diethyl ether) revealed complete reaction; the reaction mixture was evaporated at reduced pressure and directly used for the coupling without further purification.

O-Dhbt-activated resin (see Example 1)(40 mg, 0.01 mmol) was treated with *seco*-CBI dissolved in DMF (1 mL); the mixture was agitated for 1 hour at room temperature. The reaction mixture was then filtered and the resin was washed with DMF (1 mL), dichloromethane (1 mL), methanol (1 mL) and diethyl ether (2x1 mL). After thorough evaporation of excess solvent the resin was stored at -20°C.

Example 3 - Synthesis of a CBI - hexapeptide library

<u>General Procedures</u>
General procedure for acetylation/endcapping

5

After each coupling step the resin was treated with a mixture of pyridine (30%) and acetic anhydride (20%) in dichloromethane, to acetylate any free amino groups that had not been coupled to an amino acid. In this way the formation of undesirable

10   oligopeptides (carrying less than the expected number of amino acids) could be avoided.
The resin was treated with the acetylating reagent (3 mL) and the slurry was agitated at room temperature for 1 hour. The reagents were then removed by filtration and the resin was rinsed with

15   dichloromethane (2 x 5 mL) and methanol (2 x 5 mL).

*General procedure for Fmoc deprotection*
The resin was treated with a solution of piperidine in DMF (20%, 3 mL). The mixture was then agitated for 2 hours at room

20   temperature. Excess solvent was then removed by suction and the resin was rinsed with DMF (2 x 5 mL), dichloromethane (2 x 5 mL) and methanol (2 x 5 mL).

<u>Library Generation</u>

25   *Fmoc-Glu-OAll coupled to NovaSyn TG resin*
NovaSyn TG amino resin (0.345 g, load 0.29 mmol/g, 0.1 mmol) was suspended and swollen in DMF (2 mL), under agitation (1000 rpm) for 30 minutes. Excess solvent was then removed and a solution of Fmoc-Glu-OAll (0.123g, 0.3 mmol, 3 eq), TBTU (0.096 g, 0.3 mmol, 3

30   eq) and DIPEA (0.052 mL, 0.3 mmol, 3 eq) in DMF (3 mL) was added to the swollen resin. The resulting mixture was agitated at 1000 rpm at room temperature, overnight. The reaction mixture was filtered and the resin rinsed with DMF (2 x 2 mL), dichloromethane (2 x 5 mL) and methanol (2 x 5 mL).

35

The resin was then acetylated (see General procedure for acetylation) and deprotected (see General procedure for Fmoc deprotection).

HO-Gly-Fmoc coupled to P-Glu(OAll)-H
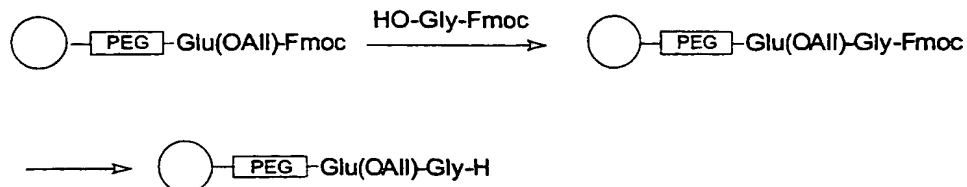
The resin was suspended in DMF (2 mL) and allowed to swell for 30
minutes at room temperature, accompanied by agitation (1000 rpm).
Excess DMF was removed and a solution of Fmoc-Gly-OH (0.089 g, 0.3
mmol, 3 eq), TBTU (0.096 g, 0.3 mmol, 3 eq) and DIPEA (0.052 mL,
0.3 mmol, 3 eq) in DMF (3 mL) was added to the resin. The mixture
was allowed to shake at room temperature for 12 hours. Excess
reagents were then removed by filtration and the resin was rinsed
with DMF (2 x 5 mL), dichloromethane (2 x 5 mL) and methanol (2 x
5 mL). The resin was then acetylated (see General procedure for
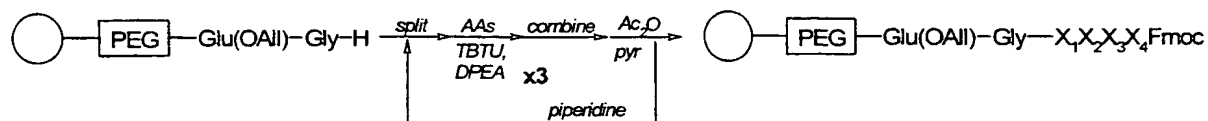acetylation) and deprotected (see General procedure for Fmoc
deprotection) to afford the resin-bound dipeptide P-Glu(OAll)-Gly-
H.

Split & Mix procedure for the resin bound hexapeptide
P-Glu(OAll)-Gly-$X_1X_2X_3X_4$-H sublibrary

The resin was suspended in 3:1 mixture of 1,2-dichloroethane (DCE)
and DMF and equally partitioned into 17 4 mL Alltech tubes. Each
tube thus contained 0.1/17 mmol = 5.88 10^{-6} mol of resin-bound
dipeptide. Excess solvent was removed *in vacuo*, and the resin was
suspended in DMF (200 mL) and agitated for 30 minutes. The 17
amino acids (1.76 10^{-5} mmol, 3 eq for each step, 7.04 10^{-5} mmol for
4 steps) were weighed into 17 vials:

1.   Fmoc-Ala-OH   22 mg
2.   Fmoc-Asn-OH   25 mg
3.   Fmoc-Asp(OtBu)-OH   29 mg

40

4.	Fmoc-Gln-OH	26 mg
5.	Fmoc-Glu(OtBu)-OH	30 mg
6.	Fmoc-Gly-OH	21 mg
7.	Fmoc-Ile-OH	25 mg
8.	Fmoc-Leu-OH	25 mg
9.	Fmoc-Lys(BOC)-OH	33 mg
10.	Fmoc-Met-OH	26 mg
11.	Fmoc-Phe-OH	27 mg
12.	Fmoc-Pro-OH	24 mg
13.	Fmoc-Ser(tBu)-OH	27 mg
14.	Fmoc-Thr(tBu)-OH	28 mg
15.	Fmoc-Trp(BOC)-OH	37 mg
16.	Fmoc-Tyr(tBu)-OH	32 mg
17.	Fmoc-Val-OH	24 mg

Each amino acid was dissolved in DMF (2 mL); an aliquot of each solution (0.5 mL, corresponding to 1.76×10^{-5} mmol, 3 eq of each amino acid) was added to the appropriate tube. TBTU (1.76×10^{-5} mmol $\times$ 17 = 2.99×10^{-4}, 96 mg) and DIPEA (1.76×10^{-5} mmol $\times$ 17 = 2.99×10^{-4}, 52 mL) were separately dissolved in DMF (1.7 mL) and each solution was evenly distributed, delivering 3 eq of each reagent, to each one of the 17 tubes.

The reaction tubes were agitated at room temperature for 12 hours, then the reagents and solvents were removed *in vacuo* and the resin was rinsed with DMF (2 x 1 mL each tube), DCM (2 x 1 mL each tube) and methanol (2 x 1 mL each tube). The resin was then suspended in 3:1 mixture of 1,2-dichloroethane and DMF and recombined. The recombined resin was acetylated (3 mL of acetylating reagent, 1 hour, room temperature) and deprotected (3 mL of 20% piperidine in DMF, 2 hours, room temperature).

The procedure was repeated 3 more times. At the end of the 4th amino acid coupling the deprotection step was not executed.

<u>Deprotection of Allyl ester</u>
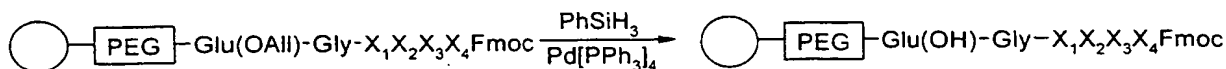
5   The resin was suspended in DCM (2 mL), phenylsilane (2.4 mmol, 24 eq, 0.29 mL) in DCM (1 mL) of was added and the mixture was shaken at room temperature for 10 minutes.

A catalytic amount of $Pd[PPh_3]_4$ (0.01 mmol, 0.1 eq, 11 mg) in DCM

10   (0.5 mL) was added and the reaction mixture was shaken for further 10 minutes.

The reagents were filtered and the resin was rinsed with DCM (2 x 5 mL) and methanol (2 x 5 mL). The procedure was repeated once

15   again and the resin was finally dried under reduced pressure.

<u>Activation of resin with 3-hydroxy-1,2,3-benzotriazin-4(3H)-one(HO-Dhbt)</u>
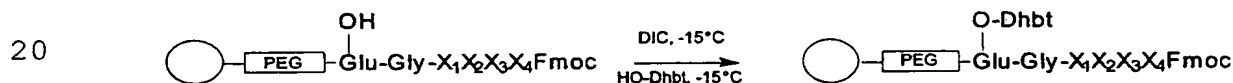
20   The resin was suspended and swollen in DMF (2 mL) for 10 minutes. Excess DMF was removed and the resin was treated with a solution of diisopropyl carbodiimide (DIC) (156 mL, 1 mmol, 10 eq) in DMF

25   (2 mL). The resulting slurry was shaken for 15 minutes at -15°C (acetone-ice bath), then washed with DMF (2 x 2 mL).

The resin was resuspended in DMF (2 mL), cooled to -10°C and treated with HO-Dhbt (163 mg 1 mmol, 10 eq). The mixture was

30   shaken for 30 minutes at -10°C, followed by 4 hours at 0°C and then allowed to stand at 0°C overnight.

The reaction mixture was filtered and the resin washed with DMF (2 x 2 mL), dichloromethane (2 x 2 mL), MeOH (2 x 2 mL) and diethyl

35   ether (2 mL). The activated resin was stored in freezer until required for use.

Coupling of O-Dhbt activated resin with *seco*-CBI, in presence of DIPEA

O-Dhbt   CBI
 | |
◯─[PEG]─Glu-Gly-X₁X₂X₃X₄Fmoc ──CBI, DPEA──▶ ◯─[PEG]─Glu-Gly-X₁X₂X₃X₄Fmoc

BOC-*seco*-CBI (see example 2) was deprotected immediately before coupling to the activated resin. BOC-*seco*-CBI (100 mg, 0.3 mmol) was dissolved in a solution of HCl in anhydrous ethyl acetate (3M, 15 mL). The mixture was cooled at 0°C for 30 minutes and then allowed to warm to room temperature over 30 minutes. TLC (4:1 = petroleum ether:diethyl ether) revealed complete reaction; the reaction mixture was evaporated under reduced pressure and stored at -20°C.

O-Dhbt-activated resin was treated with *seco*-CBI dissolved in DMF (2 mL) and the mixture was shaken for 60 minutes at room temperature. The reaction mixture was filtered and the resin was washed with DMF (2 x 2 mL), dichloromethane (2 x 2 mL), methanol (2 x 2 mL) and diethyl ether (2 x 1 mL). The resin was dried under reduced pressure and stored in a cool, dry place.

t-Butyl- and BOC- deprotection on side chains
The resin was treated with a solution of TIS/TFA in dichloromethane (2%, 2 mL). The mixture was shaken for 2 hours at room temperature. Excess reagents were removed *in vacuo* and the resin was rinsed with dichloromethane (2 x 5 mL) and methanol (2 x 5 mL).

Fmoc deprotection

CBI   CBI
 | |
◯─[PEG]─Glu-Gly-X₁X₂X₃X₄Fmoc ──piperidine──▶ ◯─[PEG]─Glu-Gly-X₁X₂X₃X₄-H
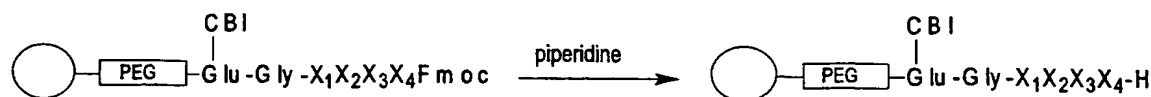
The resin was treated with a solution of piperidine in DMF(20%, 3 mL), and agitated at room temperature for 2 hours. The mixture was filtered and the resin was washed with DMF (2 x 5 mL),

dichloromethane (2 x 5 mL) and methanol (2 x 5 mL). The resin was
then dried under reduced pressure.

Screening

5   The resulting library was screened against rhodamine labelled
double stranded DNA with the sequence Label-5'-GCG TAA AAA CGC =
3'.

The sublibrary was mixed with the DNA sequence (5 pmol/mL) and
10   incubated at 37°C for 24 hours with occasional mixing. After 24
hours, the sublibrary was washed 4 times with TE buffer pH 7.6 or
PBS. To identify the beads to which most labelled DNA had bound,
agarose gel slides were prepared as follows. ~ 500 mL of 0.25% sea
plaque agarose was layered onto a clean transparent slide and
15   allowed to cool and set. The incubated beads were then mixed with
another - 500 mL of 0.25% sea plaque agarose solution and layered
onto the precoated slides, and allowed to cool and set.

The reddest beads were identified by eye under a dissecting light
20   microscope, and then retrieved by adding ~ 1 mL of water to dried
agarose slide to enable their removal using a p10 gilson pipette
with a fine tip. The removed beads were then placed into a 1 mL
Eppendorf PCR tube ready for identification.

25   ## Identification

The identification of the sequences of the most active compounds
was carried out using automated Edman degradation and reversed-
phase HPLC.

30   Pulsed liquid-phase N-terminal sequencing was performed using an
Applied Biosystems (ABI) 477A automatic protein sequencer. The
selected labelled beads were loaded onto a glass fibre disc which
had previously been pre-cycled once. The disc was placed in the
sequencer and pre-cycled once, then six cycles of Edman
35   degradation were performed (Edman, P and Begg, G (1967) Eur. J.
Biochem. 1, 80). The released phenylthiohydantion (PTH-) amino
acid derivatives were identified by reversed-phase HPLC analysis.

44

The four most active compounds were those with the following
sequences:

CBI-QGVKKK   CBI-QGLVAG
5   CBI-QGNKKA   CBI-QGQKNS

**Example 4:   Synthesis of A CBI Combinatorial Building Block: 7-
carboxy-1,2,9,9a-tetrahydrocyclopropa[c]benzo[e]indol-4-one**

10   The synthesis proceeds from a modified Stobbe
condensation/Friedel-Crafts acylation for generation of the
functionalised precursor, followed by 5-*exo-trig* aryl radical-
alkene cyclization.

15   Wadsworth-Horner-Emmons condensation of 3-bromo-benzaldehyde with
the Sargent phosphonate predominantly provides the E-isomer, which
in turn undergoes acid-catalyzed deprotection and Friedel-Crafts
acylation (Scheme 6).

Scheme 6
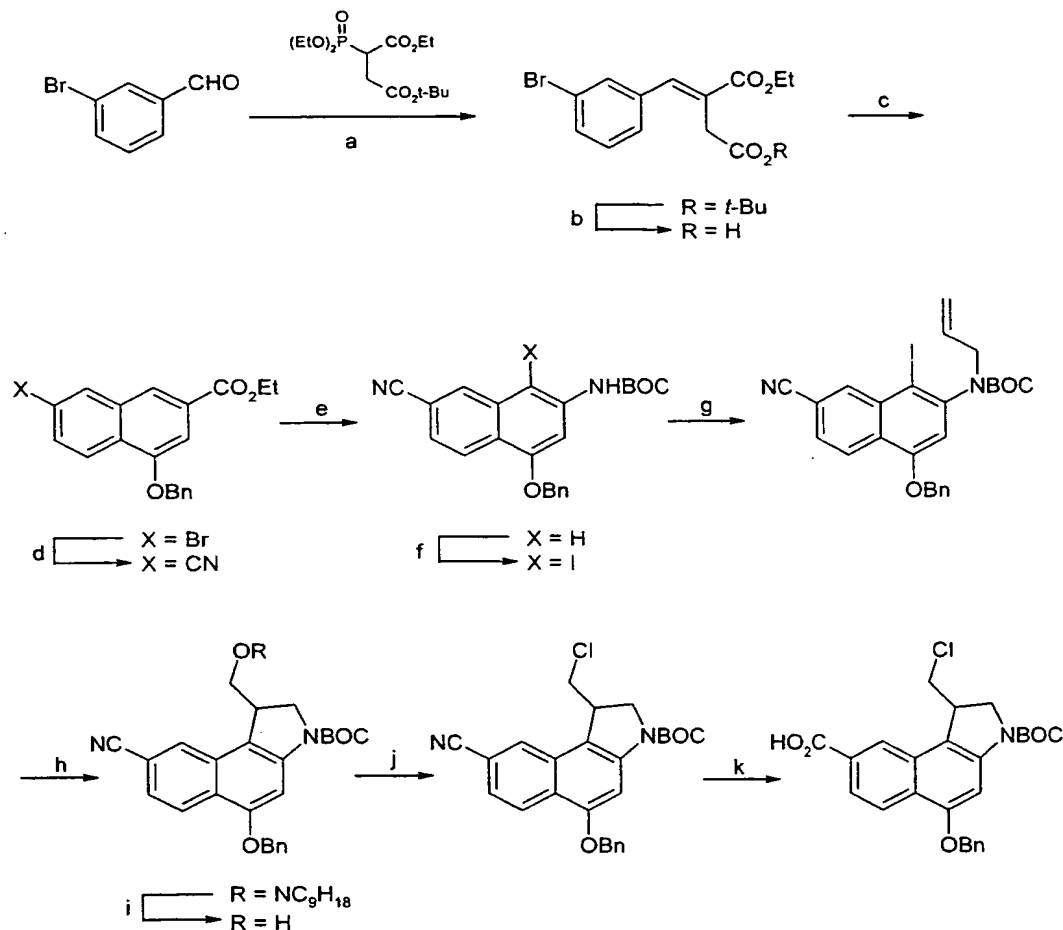
Reagents and conditions: **a**: NaH, Sargent phosphonate; **b**: TFA; **c**: 1) Ac_2O-KOAc; 2) K_2CO_3; 3) BnBr, K_2CO_3; **d**: CuCN; **e**: 1) LiOH; 2) DPPA, t-BuOH; **f**: NIS; **g**: allyl Br, NaH; **h**: Bu_3SnH, TEMPO; **i**: Zn-HOAc; **j**: Ph_3P-CCl_4; **k**: NaOH

Aromatic nucleophilic substitution, ester hydrolysis and Curtius rearrangement effected by treatment with DPPA are followed by regioselective C4 iodination and N-alkylation with allyl bromide. The aryl radical-alkene cyclization by means of TEMPO as radical trap, as described in Boger synthesis of CBI, provides the tricyclic system that, after conversion to the primary chloride and base-catalyzed hydrolysis of the cyano group, should give the desired building block for a combinatorial library.

Example 5: Screening of CBI on bead against DNA sequence

The CBI on bead synthesised in example 1 was screened against rhodamine labelled double stranded DNA with the sequence:

5   Label-5'-GCG TAA AAA CGC-3'.

For comparison, indoline was added to O-Dhbt-activated TG Carboxy resin (as prepared in example 1) to yield a comparative resin that does not covalently interact with DNA.

10
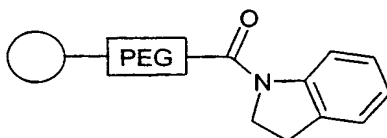
The resin alone was also tested to provide a background measurement. The screening protocol used was:

15

1.   Weigh out approximately 1 mg of resin into an Eppendorf.
2.   Incubate the resin with annealed rhodamine labelled double strand DNA for 24 hours at 37°C.
3.   After 24 h incubation wash the resin 4 times and re-suspend
20   beads in 50 mL of TE (EDTA and Tris buffer) or PBS (phosphate buffered saline).
4.   Resin was transferred to a black 96 well plate with transparent base, fluorescence was measured from below at 635 nm using a Tecan Spectrofluor (590 nm excitation, 635 nm
25   emission).

Resin	Relative Fluorescence Units
TG Carboxy Resin	892
Indoline Bound to Resin	625
CBI Bound to Resin	2947

30

Only a small amount of DNA binding was observed with the acid resin and with the resin loaded with an inactive indoline CBI mimic. An almost five fold differential was observed between the
35   CBI resin and Indoline resin demonstrating DNA binding by the resin bound CBI. It is anticipated that a higher concentration of DNA would lead to a higher differential between the resins as it

47

is possible that the CBI-resin is not saturated with DNA at this
concentration.

Example 6: Synthesis of a CBI - PBD - hexapeptide library

5

General procedure for acetylation/endcapping

After any coupling step the resin was treated with a mixture of
pyridine (30%) and acetic anhydride (20%) in dichloromethane, to
10   acetylate any free amine that has not been coupled to an amino
acid. The formation of unexpected oligopeptides (carrying less
than the expected number of aminoacids) is avoided.

The resin was treated with 3 mL of acetylating reagent and the
15   slurry was agitated at room temperature for 1 hour. The reagents
were then removed by filtration and the resin was rinsed with
dichloromethane (2 x 5 mL) and methanol (2 x 5 mL).

General procedure for Fmoc deprotection

20   The resin was treated with 3 mL of a solution of piperidine (20%)
in DMF. The mixture was then agitated for 2 hours at room
temperature. The solvent was filtered off and the resin was
rinsed with DMF (2 x 5 mL), dichloromethane (2 x 5 mL) and
25   methanol (2 x 5 mL).

Fmoc-Glu-OAll coupled to NovaSyn TG resin

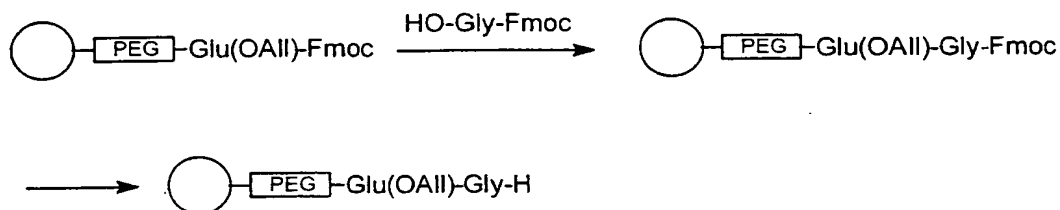30   ⬤─[PEG]─NH₂ + Fmoc-Glu-OAll ⟶ ⬤─[PEG]─Glu(OAll)-Fmoc ⟶ ⬤─[PEG]─Glu(OAll)-H

NovaSyn TG amino resin 130 μ (0.345 g, load 0.29 mmol/g, 0.1 mmol)
was suspended ad swollen into 2 mL of DMF, under agitation (1000
rpm) for 30 minutes. The solvent was then removed and a solution
35   of Fmoc-Glu-OAll (0.123g, 0.3 mmol, 3 eq), TBTU (0.096g, 0.3 mmol,
3 eq) and DIPEA (0.052 mL, 0.3 mmol, 3 eq) in 3 mL of DMF was
added to the swollen resin. The resulting mixture was agitated at
1000 rpm, room temperature, overnight. The reaction mixture was

filtered and the resin rinsed with DMF (2 x 2 mL), dichloromethane
(2 x 5 mL) and methanol (2 x 5 mL).

5   The resin was then acetylated (see General procedure for
acetylation) and deprotected (see General procedure for Fmoc
deprotection).

HO-Gly-Fmoc coupled to P-Glu(OAll)-H

10

The resin was suspended in 2 mL of DMF and was allowed to swell
for 30 minutes at room temperature, whilst being agitated (1000
15   rpm). DMF was removed and a solution of Fmoc-Gly-OH (0.089 g, 0.3
mmol, 3 eq), TBTU (0.096 g, 0.3 mmol, 3 eq) and DIPEA (0.052 mL,
0.3 mmol, 3 eq) in 3 mL of DMF was added to the resin. The mixture
was shaken at room temperature for 12 hours. The reagents were
then removed by filtration and the resin was rinsed with DMF (2
20   x5 mL), dichloromethane (2 x 5 mL) and methanol (2 x 5 mL). The
resin was then acetylated (see General procedure for acetylation)
and deprotected (see General procedure for Fmoc deprotection) to
afford the resin-bound dipeptide P-Glu(OAll)-Gly-H.

25   Split & Mix procedure for the resin bound hexapeptide P-Glu(OAll)-
Gly-$X_1X_2X_3X_4$-H sublibrary
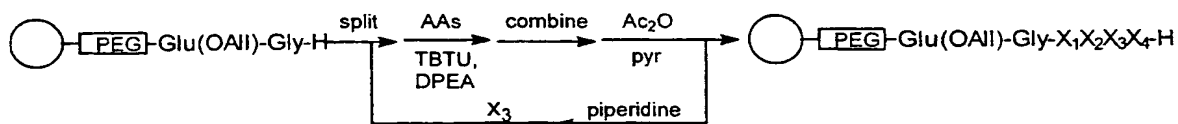
30

The resin was suspended in 3:1 mixture of 1,2-dichloroethane and
DMF and equally partitioned into 17 4 mL Alltech tubes. Each tube
thus contained $0.1/17$ mmol = $5.88 \; 10^{-6}$ mol of resin-bound
dipeptide. Solvent was removed in vacuo, and the resin was

49

suspended in 200 mL of DMF and agitated for 30 minutes. The 17 aminoacids ($1.76 \ 10^{-5}$ mmol, 3 eq for each step, $7.04 \ 10^{-5}$ mmol for 4 steps) were weighed into 17 vials:

|			
5	1.	Fmoc-Ala-OH	22 mg
	2.	Fmoc-Asn-OH	25 mg
	3.	Fmoc-Asp(OtBu)-OH	29 mg
	4.	Fmoc-Gln-OH	26 mg
	5.	Fmoc-Glu(OtBu)-OH	30 mg
10	6.	Fmoc-Gly-OH	21 mg
	7.	Fmoc-Ile-OH	25 mg
	8.	Fmoc-Leu-OH	25 mg
	9.	Fmoc-Lys(BOC)-OH	33 mg
	10.	Fmoc-Met-OH	26 mg
15	11.	Fmoc-Phe-OH	27 mg
	12.	Fmoc-Pro-OH	24 mg
	13.	Fmoc-Ser(tBu)-OH	27 mg
	14.	Fmoc-Thr(tBu)-OH	28 mg
	15.	Fmoc-Trp(BOC)-OH	37 mg
20	16.	Fmoc-Tyr(tBu)-OH	32 mg
	17.	Fmoc-Val-OH	24 mg

Each amino acid was dissolved in 2 mL of DMF; 0.5 mL of this solution (corresponding to $1.76 \ 10^{-5}$ mmol, 3 eq of each amino acid) was added to the appropriate tube. TBTU ($1.76 \ 10^{-5}$ mmol x 17 = $2.99 \ 10^{-4}$, 96 mg) and DIPEA ($1.76 \ 10^{-5}$ mmol x 17 = $2.99 \ 10^{-4}$, 52 mL) were separately dissolved in 1.7 mL of DMF and each solution was evenly partitioned, addressing 3 eq of each reagent, to each one of the 17 tubes.

The reaction tubes were agitated at room temperature for 12 hours, then the reagents and solvents were removed *in vacuo* and the resin was rinsed with DMF (2 x 1 mL each tube), DCM (2 x 1 mL each tube) and methanol (2 x 1 mL each tube). The resin was then suspended in 3:1 mixture of 1,2-dichloroethane and DMF and recombined. The recombined resin was acetylated (3 mL of acetylating reagent, 1 hour, room temperature) and deprotected (3 mL of 20% piperidine in DMF, 2 hours, room temperature).

The procedure was repeated 3 more times to afford the deprotected resin-bound hexapeptide $G(OAll)EX_1X_2X_3X_4H$.

Coupling of Fmoc(Alloc)Lys-OH to resin-bound hexapeptide
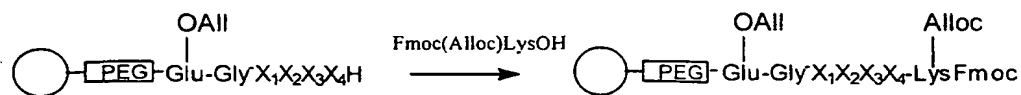
The resin was suspended in 2 mL of DMF and was allowed to swell for 10 minutes at room temperature, whilst being agitated (1000 rpm). DMF was removed and a solution of Fmoc(Alloc)Lys-OH (0.135g, 0.3 mmol, 3 eq), TBTU (0.096g, 0.3 mmol, 3 eq) and DIPEA (0.052 mL, 0.3 mmol, 3 eq) in 3 mL of DMF was added to the resin. The mixture was shaken at room temperature for 12 hours. The reagents were then removed by filtration and the resin was rinsed with DMF (2 x 5 mL), dichloromethane (2 x 5 mL) and methanol (2 x 5 mL). The resin was then acetylated (see General procedure for acetylation) to afford the resin-bound heptapeptide.

Deprotection of Allyl ester
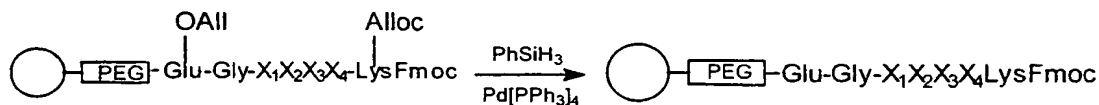
The resin was suspended in 2 mL of DCM. Phenylsilane (2.4 mmol, 24 eq, 0.29 mL) in 1 mL of DCM was added and the mixture was shaken at room temperature for 10 minutes. Pd[PPH$_3$]$_4$ (0.01 mmol, 0.1 eq, 11 mg) in 0.5 mL of DCM was added and the reaction mixture was shaken for further 10 minutes.

The reagents were filtered and the resin was rinsed with DCM (2 x 5 mL) and methanol (2 x 5 mL), then the procedure was repeated once again. The resin was finally dried under reduced pressure.

<u>Coupling of N10-Fmoc protected PBD to lysine residue of the resin-bound heptapeptide</u>
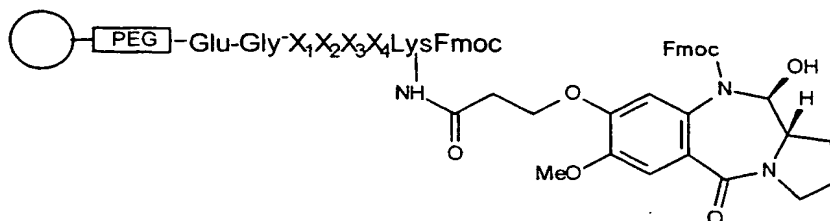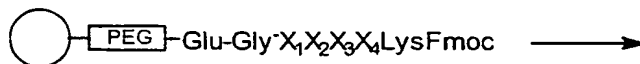
The resin was suspended in 2 mL of DMF and agitated for 10 minutes
to allow swelling. DMF was then removed. A solution of N10-Fmoc-
PBD acid (0.556 g, 1 mmol, 10 eq.), TBTU (0.353 g, 1.1 mmol, 1.1
eq.) and DIPEA (0.19 mL, 1.1 mmol, 1.1 eq.) in 3 mL of DMF was
stirred for 30 minutes, and added to the resin. The mixture was
agitated at room temperature for 12 hours, then the reagents were
removed under reduced pressure and the resin was rinsed with DMF
(2 x 5 mL), dichloromethane (2 x 5 mL), methanol (2 x 5 mL) and
diethyl ether (1 x 2 mL).

<u>Activation of resin with 3-hydroxy-1,2,3-benzotriazin-4(3H)-one
(HO-Dhbt)</u>
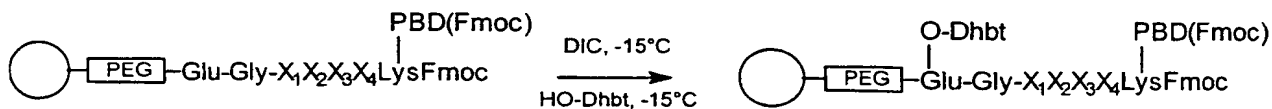
The resin was suspended and swollen in 2 mL of DMF for 10 minutes.
DMF was removed and the resin was treated with 156 mL (1 mmol,
10 eq) of diisopropyl carbodiimide (DIC) in 2 mL of DMF. The
resulting slurry was shaken for 15 minutes at −15°C (acetone-ice
bath), then washed with DMF (2 x 2 mL).

The resin was re-suspended in 2 mL of DMF, cooled at -10°C and treated with 163 mg (1 mmol, 10 eq) of HO-Dhbt. The mixture was shaken for 30 minutes at -10°C, for 4 hours at 0°C and then allowed to stand at 0°C overnight.

The reaction mixture was filtered and the resin washed with DMF (2 x 2 mL), dichloromethane (2 x 2 mL), MeOH (2 x 2 mL) and diethyl ether (2 mL). The activated resin was stored in freezer until ready for use.

Coupling of O-Dhbt activated resin with *seco*-CBI, in presence of DIPEA
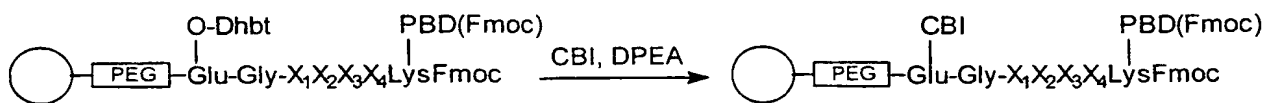
BOC-*seco*-CBI was deprotected immediately before coupling to the activated resin. 100 mg (0.3 mmol) of BOC-*seco*-CBI was dissolved in 15 mL of 3M HCl solution in anhydrous ethyl acetate. The mixture was cooled at 0°C for 30 minutes and then allowed to warm to room temperature for further 30 minutes. TLC (4:1 = petroleum ether:diethyl ether) revealed reaction completion; the reaction mixture was evaporated at reduced pressure and stored at -20°C.
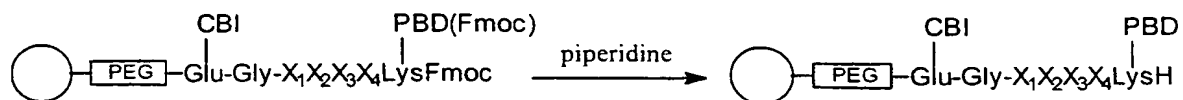
O-Dhbt-activated resin was treated with *seco*-CBI dissolved in 2 mL of DMF and the mixture was shaken for 60 minutes at room temperature. At the end the reaction mixture was filtered and the resin was washed with DMF (2 x 2 mL), dichloromethane (2 x 2 mL), methanol (2 x 2 mL) and diethyl ether (2 x 1 mL). The resin was dried under reduced pressure and stored in a cool, dry place.

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t-Butyl- and BOC- deprotection on side chains

The resin was treated with a solution of TIS/TFA 2% in dichloromethane (2 mL). The mixture was shaken for 2 hours at room temperature. The reagents were removed *in vacuo* and the resin was rinsed with dichloromethane (2 x 5 mL) and methanol (2 x 5 mL).

Fmoc deprotection

$$
\underset{\text{PEG}}{\bigcirc}-\underset{\begin{subarray}{c}\text{CBI}\end{subarray}}{\boxed{\text{PEG}}}-\text{Glu-Gly-}X_1X_2X_3X_4\underset{\begin{subarray}{c}\text{PBD(Fmoc)}\end{subarray}}{\text{Lys}}\text{Fmoc} \xrightarrow{\text{piperidine}} \underset{\text{PEG}}{\bigcirc}-\boxed{\text{PEG}}-\text{Glu-Gly-}X_1X_2X_3X_4\text{Lys}H
$$

CBI — PBD(Fmoc) → CBI — PBD

The resin was treated with 3 mL of a solution of piperidine (20%) in DMF, and agitated at room temperature for 2 hours. At the end the mixture was filtered and the resin was washed with DMF (2 x 5 mL), dichloromethane (2 x 5 mL) and methanol (2 x 5 mL). The resin was then dried under reduced pressure.